

# Cell Ontologies, Annotation & Metadata Session @ HCA on June 28, 2022

[https://cns-iu.github.io/workshops/2022-06-27 human cell atlas/](https://cns-iu.github.io/workshops/2022-06-27_human_cell_atlas/)

[https://bit.ly/HCA\\_Cell\\_ont](https://bit.ly/HCA_Cell_ont)



# Overview



## Time & Date

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June 28, 2022 from 10:10-11:40 GMT+1 (1 hour, 30 mins)

Event webpage: <https://www.humancellatlas.org/hcameetings>

## Location

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Human Cell Atlas General Meeting

Vienna, Austria

(For virtual attendance, see <https://www.humancellatlas.org/hcameetings>)

## Goals

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The goal of the breakout session is to discuss challenges and propose solutions to the development and use of ontologies for FAIR sharing and integration of human cell atlas (HCA) data for atlas construction and usage (e.g., in the **Human Reference Atlas**).

[https://cns-iu.github.io/workshops/2022-06-27\\_human\\_cell\\_atlas/](https://cns-iu.github.io/workshops/2022-06-27_human_cell_atlas/)



## Organizers

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Katy Börner  
Indiana University, USA  
[katy@indiana.edu](mailto:katy@indiana.edu)



David Osumi-Sutherland  
EBI, UK  
[davidos@ebi.ac.uk](mailto:davidos@ebi.ac.uk)

## Key Speakers

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Evan Biederstedt  
Harvard Medical School, USA



Melissa Clarkson  
University of Kentucky College of  
Medicine, USA



Bruce W. Herr II  
Indiana University, USA  
[Slides](#) | [Video](#)



Jason Hilton  
Stanford University, USA



Wei Kheng Teh  
Archival Infrastructure and  
Technology, EBI, UK



Angela Pisco  
CZI BioHub, USA



Fabian Theis  
Helmholtz-Muenchen, Germany



## Summary

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Standardizing the way we annotate samples and analysis metadata but also anatomical structures, cell types, and biomarkers is a key component of making HCA data Findable, Accessible, Interoperable and Re-usable (FAIR) and ultimately to integrating it into coherent atlases such as the **Human Reference Atlas (HRA)**. Ontologies, combined with standard annotation schemas, aid this process by providing standard terms for annotation and mechanisms for grouping terms in biologically meaningful ways, for example, grouping cell types by location or function. This session will discuss key challenges we face in achieving these aims and the opportunities that will be opened up by achieving them:

1. How can we **extend and improve ontologies** as our knowledge grows leveraging expert input, experimental data and feedback from different atlasing efforts?
2. How can we **make ontology annotation easy**, efficient, and accurate while leaving room for revising and adding to existing ontologies?
3. How can we enable downstream users to take advantage of ontology structure and content in **analysis, visualization and machine learning pipelines/applications**?
4. How can improved annotation with ontologies and the use of linked open data (LOD) help us to interlink atlas data and from multiple consortia and **construct more integrated, coherent, and queryable atlases**?



# Agenda

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## Welcome and Introduction | June 28, 2022: 10.10 – 10.15 (GMT+1)

- Introduction of Workshop Goals by Session Organizers

## Flash Talks | June 28, 2022: 10.15 – 10.45 (GMT+1)

- Six 5 minute flash talks on one of the challenges/opportunities, with participants talking about how their work can help &/or challenge they need, help solving

## Breakout Introduction | June 28, 2022: 10.45 – 11.00 (GMT+1)

- Each group must nominate a scribe and a chair. Breakout groups will fill out structured forms for use in report back

## Breakouts | June 28, 2022: 11.00 – 11.40 (GMT+1)

- Four breakouts, one on each of the four challenges/opportunities. Self assorting. Each group must nominate a scribe and a chair. Breakout groups will fill out structured form for use in report-back in the main session



## Relevant Papers

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- Börner, Katy, Sarah A Teichmann, Ellen M Quardokus, et al. 2021. **"Anatomical structures, cell types and biomarkers of the Human Reference Atlas"** . *Nature Cell Biology* 23: 1117-1128. doi: 10.1038/s41556-021-00788-6.
- Börner, Katy, Andreas Bueckle, Bruce W. Herr II, et al. 2021. **"Tissue Registration and Exploration User Interfaces in support of a Human Reference Atlas"**. *bioRxiv* doi: 10.1101/2021.12.30.474265.
- Osumi-Sutherland, David, Chuan Xu, Maria Keays, Adam P. Levine, Peter V. Kharchenko, Aviv Regev, Ed Lein, and Sarah A. Teichmann. 2021. "Cell Type Ontologies of the Human Cell Atlas." *Nature Cell Biology* 23 (11): 1129–35.
- M. D. Wilkinson et al., "The FAIR Guiding Principles for scientific data management and stewardship," *Sci Data*, vol. 3, p. 160018, Mar. 2016, doi: 10.1038/sdata.2016.18

Please share others via <https://asct-b.slack.com>



**Welcome**



# Making cell type data findable with ontology annotation

## COVID-19 Cell Atlas



Source	Term used in data annotation
Madisoorn et al., 2019 PMID:31892341	Alveolar_Type1
Lukasen et al., 2020 DOI:10.15252/emboj.20105114	AT1
Vieira Braga et al., 2019 PMID:31209336	Type_1_alveolar
Travaglini et al. 2020 DOI:10.1038/s41586-020-2922-4	Alveolar Epithelial Type 1

### type I pneumocyte

[http://purl.obolibrary.org/obo/CL\\_0002062](http://purl.obolibrary.org/obo/CL_0002062) [Copy](#)

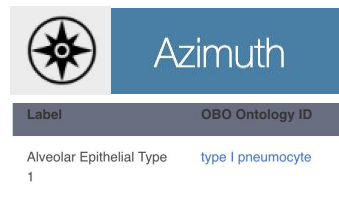
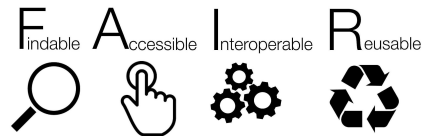
A type I pneumocyte is a flattened, branched pneumocyte that covers more than 98% of the alveolar surface. This large cell has thin (50-100 nm) cytoplasmic extensions to form the air-blood barrier essential for normal gas exchange. [ <http://www.ncbi.nlm.nih.gov/pubmed/20054144> GOC:tfm <http://www.copewithcytokines.de> ]

**Synonyms:** [pulmonary alveolar type I cell](#) [small alveolar cells](#) [ATI](#)

[type I alveolar epithelial cells](#) [squamous alveolar cell](#) [membranous pneumocytes](#)

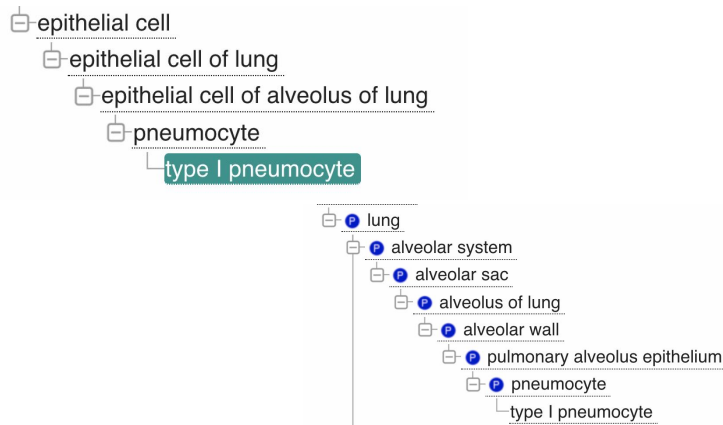
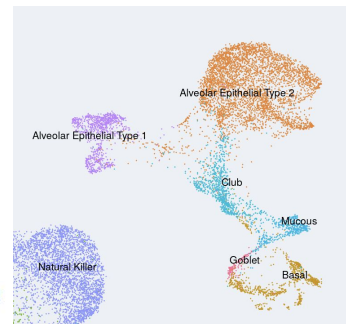
[type I alveolar cells](#) [type 1 alveolar epithelial cells](#) [type 1 pneumocyte](#)

[squamous alveolar lining cell](#) [lung type 1 cells](#) [AT1](#)



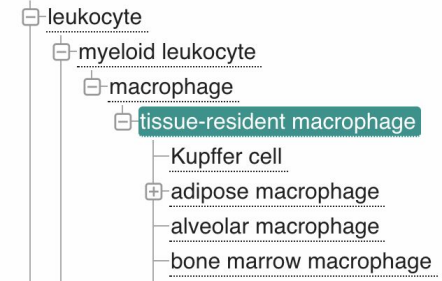
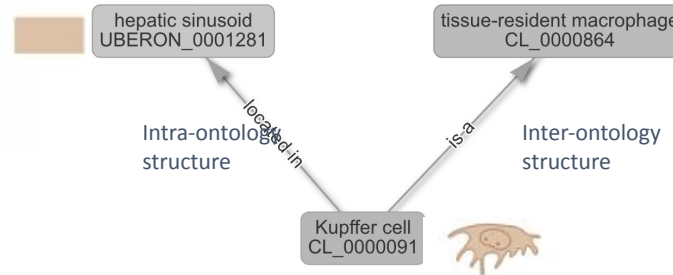
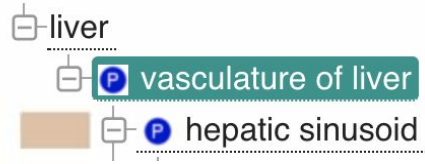
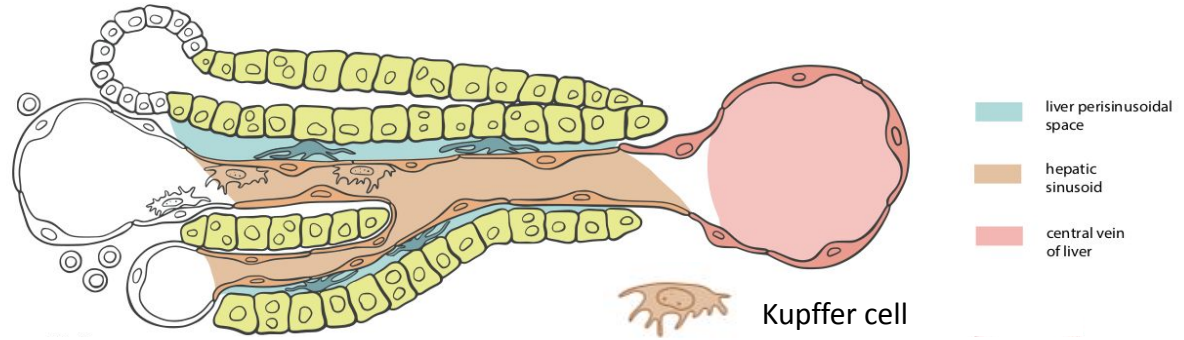
Reference Dataset(s): [Travaglini et al, Nature 2020](#)

Demo Dataset(s): [Vieira-Braga et al, Nature Medicine 2019 \[Seurat Object\]](#)



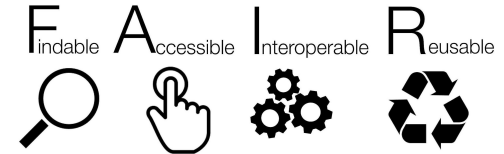


Ontology structure allows  
grouping of related content  
– making it more  
*Findable and Interoperable*



## Queries:

- Find all scRNAseq datasets with data on:
  - tissue resident macrophages
  - cells in the liver





# Discussion Topics

1. How can we **extend and improve ontologies** as our knowledge grows leveraging expert input, experimental data and feedback from different atlasing efforts?

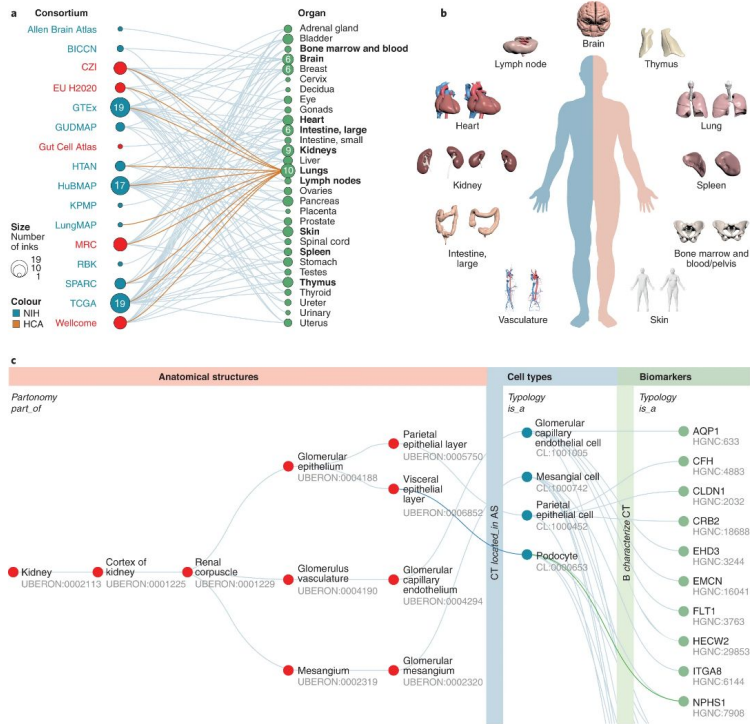
We're hiring ontology editors <https://www.embl.org/jobs/position/EBI02008>

2. How can we **make ontology annotation, following standard schemas easy**, efficient, and accurate while leaving room for revising and adding to existing ontologies?
3. How can we enable downstream users to take advantage of ontology structure and content in **analysis, visualization and machine learning pipelines/applications**?
4. How can improved annotation with ontologies and the use of linked open data (LOD) help us to interlink atlas data and from multiple consortia and **construct more integrated, coherent, and queryable atlases**?

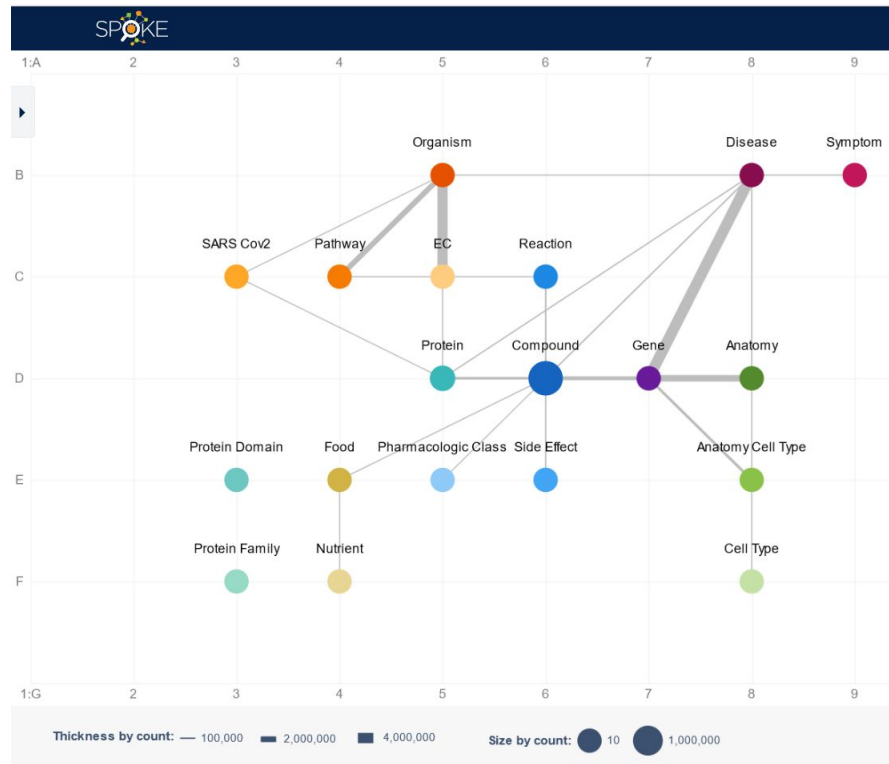
**REMINDER FOR SPEAKERS – 5' ONLY - EMPHASISE A PROBLEM/SOLUTION**



# Katy's Welcome



<https://hubmapconsortium.github.io/ccf/>  
<https://www.nature.com/articles/s41556-021-00788-6>



<https://cns-iu.github.io/spoke-vis/home>  
<https://onlinelibrary.wiley.com/doi/10.1002/aaai.12037>



# Question: How to **bidirectionally** link ontologies to 1/2/3D references

**HuBMAP** CCF EXPLORATION LOGIN

Sex: **Both** Age: **1-110** BMI: **13-83**

Search anatomical structures...

body

362

brain

0

lymph node

30

eye

0

fallopian tube

0

heart

51

atrioventricular junction

0

epicardium

0

Search cell types...

cell

362

absorptive

47

adventitial stromal cell

57

afferent neuron cell

4

airway deuterosomal cell

11

alveolar type 1 fibroblast

11

alveolar type 2 fibroblast

11

Blood Vasculature 2

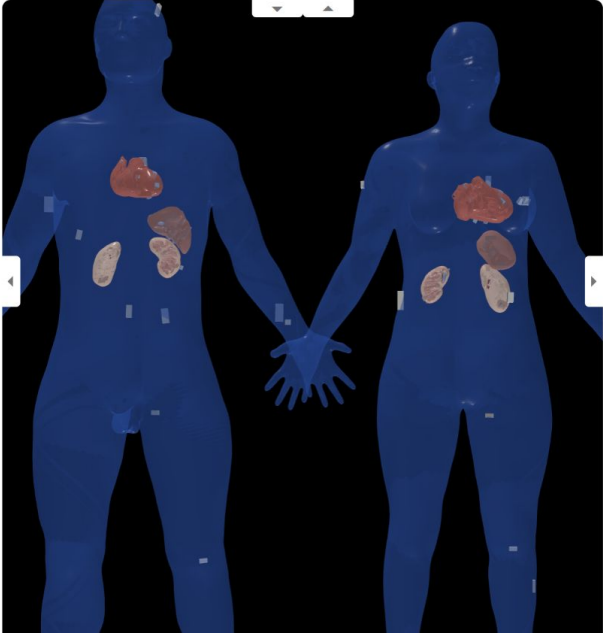
Brain 0

Eye, L 0

Eye, R 0

Fallopian Tube, L 0

Fallopian Tube, R 0



body | cell

9 Tissue Data Providers

138 Donors

362 Tissue Blocks

595 Tissue Sections

1236 Tissue Datasets

☐ **Apical Septum Female**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Basal Right Ventricle Free Wall Female**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Basal Septum Left Ventricle Female**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Basal Septum Left Ventricle Male**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Left Ventricle Apex Female**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Left Ventricle Apex Male**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Middle Anterior Left Ventricle Female**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

☐ **Middle Anterior Left Ventricle Male**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

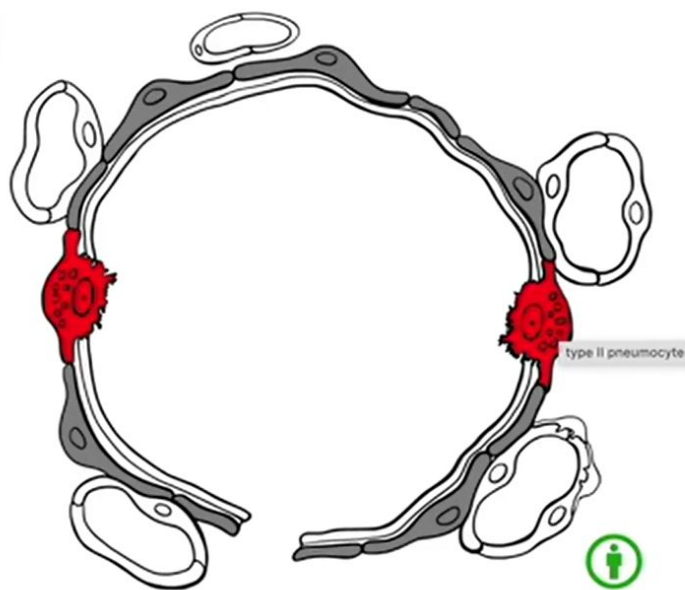
☐ **Middle Lateral Left Ventricle Female**  
Entered 3/16/2021, Peter Hanna, SPARC/UCLA

<https://portal.hubmapconsortium.org/ccf-eui>





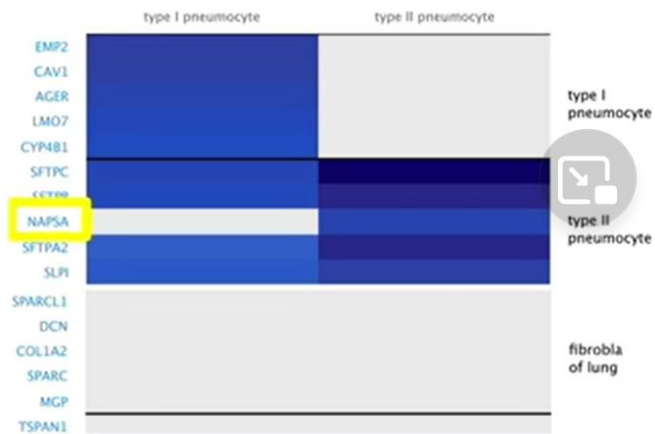
## NAPSA marker gene in type II pneumocytes



### Cell type marker genes [Download](#)

Median expression (CPM)

0.1 1 10 100 1k 10k 100k 1M





## Welcome to the Kidney Tissue Atlas Explorer

Search by marker gene, cell type, or data type to view summary data visualizations across the various KPMP 'omics' technologies.

### Search



### Select a data type

DATA TYPE	HEALTHY REFERENCE	CKD	AKI
Single-nucleus RNA-seq (snRNA-seq)*	13	10	6
Single-cell RNA-seq (scRNA-seq)*	20	15	12
Regional transcriptomics (LMD RNA-seq)	9	22	5

\* Additional information available in [cellxgene](#)

### Select a cell type

Glomerulus / Renal Corpuscle

Tubules

Interstitial

Vessels

Glomerulus / Renal Corpuscle

Glomerular Parietal Epithelium

Parietal Epithelial Cell

Glomerular Visceral Epithelium

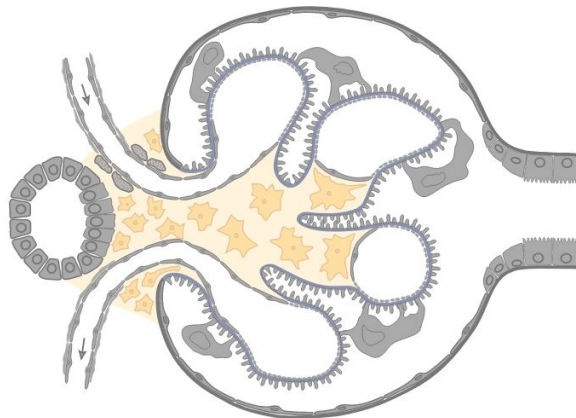
Visceral Epithelial Cell

Glomerular Capillary Endothelium

Glomerular Capillary Endothelial Cell

Glomerular Mesangium

Mesangial Cell



<https://atlas.kpmp.org/explorer/>



# HuBMAP + HPA - Hacking the Human Body

Segment multi-organ functional tissue units

\$60,000

Prize Money



HuBMAP + HPA · 84 teams · 3 months to go (3 months to go until merger deadline)

[Overview](#) [Data](#) [Code](#) [Discussion](#) [Leaderboard](#) [Rules](#)

## Overview

### Description

### Supervised ML Evaluation

### Timeline

### Prizes

### Judges Prize

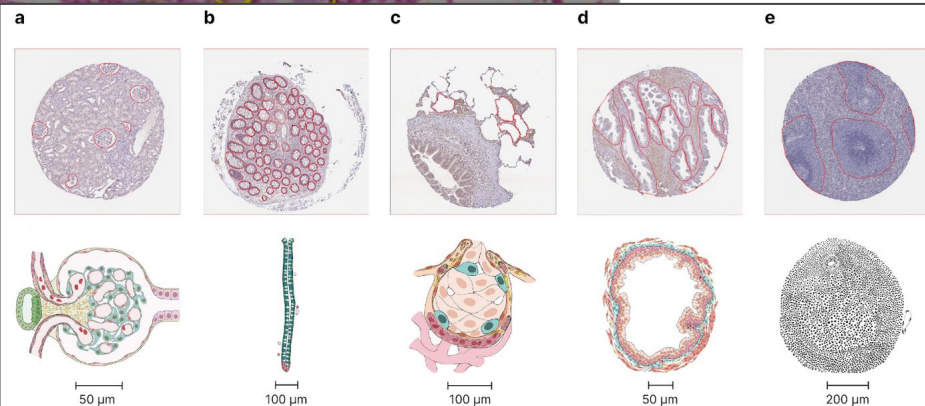
### Code Requirements

### Organizers & Sponsors

When you think of “life hacks,” normally you’d imagine productivity hacks. But what if you could “hack” your body at a molecular level? It may be possible! Researchers need to understand the trillions of cells that make up the human body. A better understanding could lead to longer lives.

A previous [Kaggle competition](#) aimed to annotate cell population function, also called functional tissue units (FTUs). Manually annotating FTUs (e.g., glomeruli in kidney or alveoli in the lung) is a time-consuming process. In the average kidney, there are over 1 million glomeruli FTUs. While there are existing cell and FTU segmentation methods, we want to push the boundaries by building algorithms that generalize across different organs and are robust across different dataset differences.

The [Human BioMolecular Atlas Program](#) (HuBMAP) is working to create a [Human Reference Atlas](#) at the cellular level. Sponsored by the National Institutes of Health (NIH), HuBMAP and Indiana University’s Cyberinfrastructure for Network Science Center (CNS) have partnered with institutions across the globe for this endeavor. A major partner is the [Human Protein Atlas](#) (HPA), a Swedish research program aiming to map the protein expression in human cells, tissues, and organs, funded by the Knut and Alice Wallenberg Foundation.





# Flash Talks



# Flash Talks by Experts

— — —

Melissa Clarkson, UKY, USA

Wei Kheng Teh, EBI, UK

Jason Hilton, Stanford U, USA

Evan Biederstedt, Harvard Medical School, USA

Angela Pisco, CZI BioHub, USA

Fabian Theis, Helmholtz-Muenchen, Germany

Bruce W. Herr II, Indiana University, USA

## **Other experts in the room**

Tony Burdett, EBI, UK

Paulo Czarnewski, Scilifelab, SE



**Melissa Clarkson**



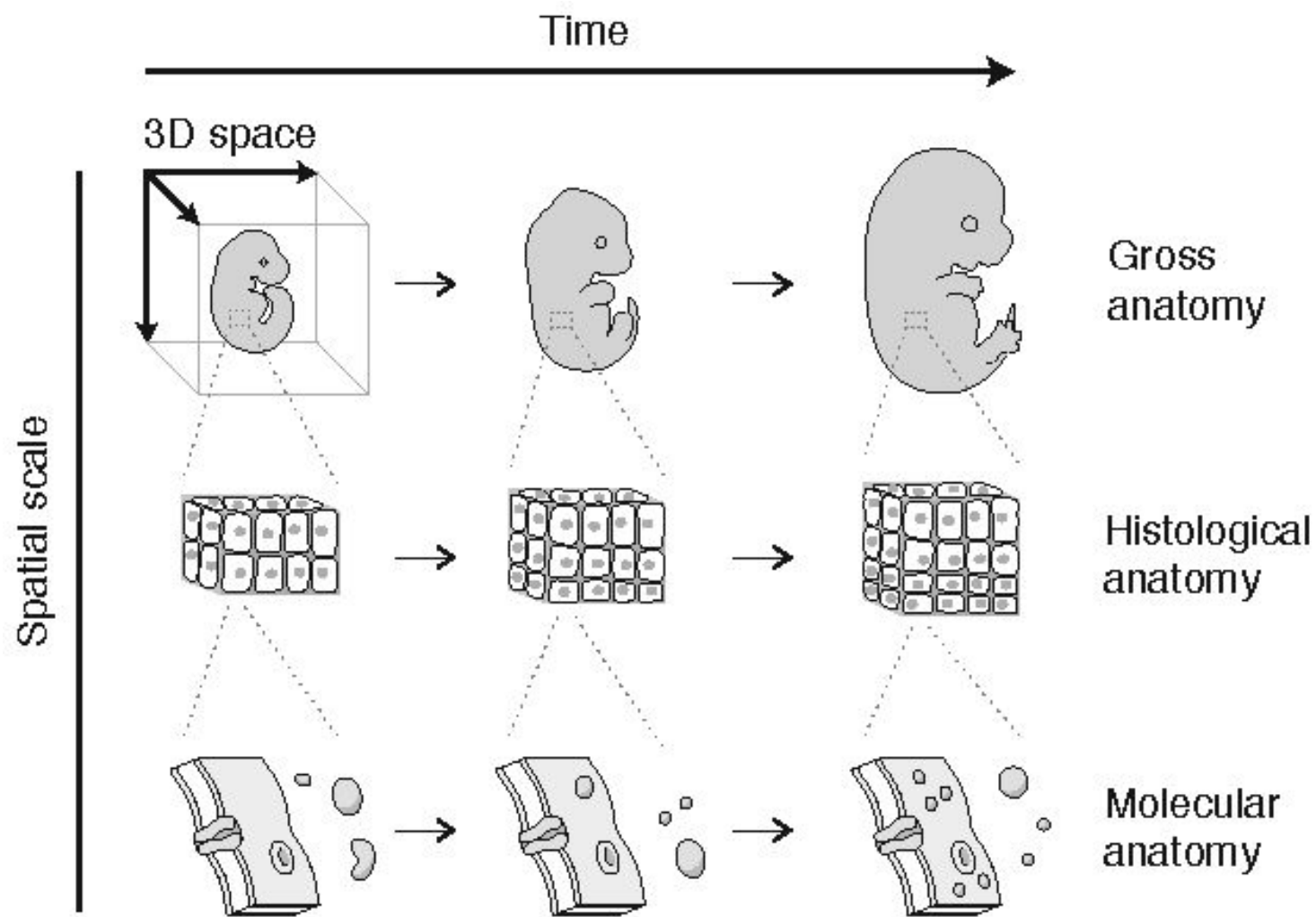
Melissa Clarkson, UKY, USA

— — —

Representing human anatomy at the scale of organs to  
tissues —

**The Foundational Model of Human Anatomy project**







# The Foundational Model of Anatomy (FMA) is a reference ontology for adult canonical anatomy

- A project of the Structural Informatics Group at the University of Washington
- Modeled in OWL
- Over 100,000 anatomical structures represented as classes
- Over 100 types of relations among classes



The FMA has a number of issues that affect its ability to continue to serve as a knowledge base

- Variations in modeling schemes for similar anatomy in different parts of the body
- Incomplete content
- Not easy to understand meaning of some classes



I am beginning a project to develop a derivative of the FMA that will eventually replace the FMA

## **Foundational Model of Human Anatomy (FMHA)**

Development strategy will improve:

- consistency of modeling
- completeness
- clarity



Consistency of modeling will be improved by using patterns

*Example of inconsistencies in the FMA...*

Question:

How is a muscle related to the bone it attaches to?



# Consistency of modeling will be improved by using patterns

## *Example of inconsistencies in the FMA*

<b>Subject superclass</b>	<b>Relation</b>	<b>Object superclass</b>	<b>Number of axioms</b>
Tendon	attaches to	Zone of bone organ	207
Zone of muscle organ	has insertion	Zone of bone organ	60
Muscle organ	has insertion	Zone of bone organ	36
Muscle organ	attaches to	Zone of bone organ	11



Consistency of modeling will be improved by using patterns

**Muscle organ**

↓ *has constitutional part*

**Tendon**

↓ *attaches to*

**Zone of bone organ**

↓ *regional part of*

**Bone organ**



The vision:

The Foundational Model of Human Anatomy (FMHA) ontology will be a computable representation of human anatomy, linked to graphics with computer-readable semantics — creating an “illustrated ontology”.



**Wei Kheng Teh**



# Wei Kheng Teh, EBI, UK

— — —



HUMAN CELL ATLAS  
**DATA PORTAL**

<https://data.humancellatlas.org/>

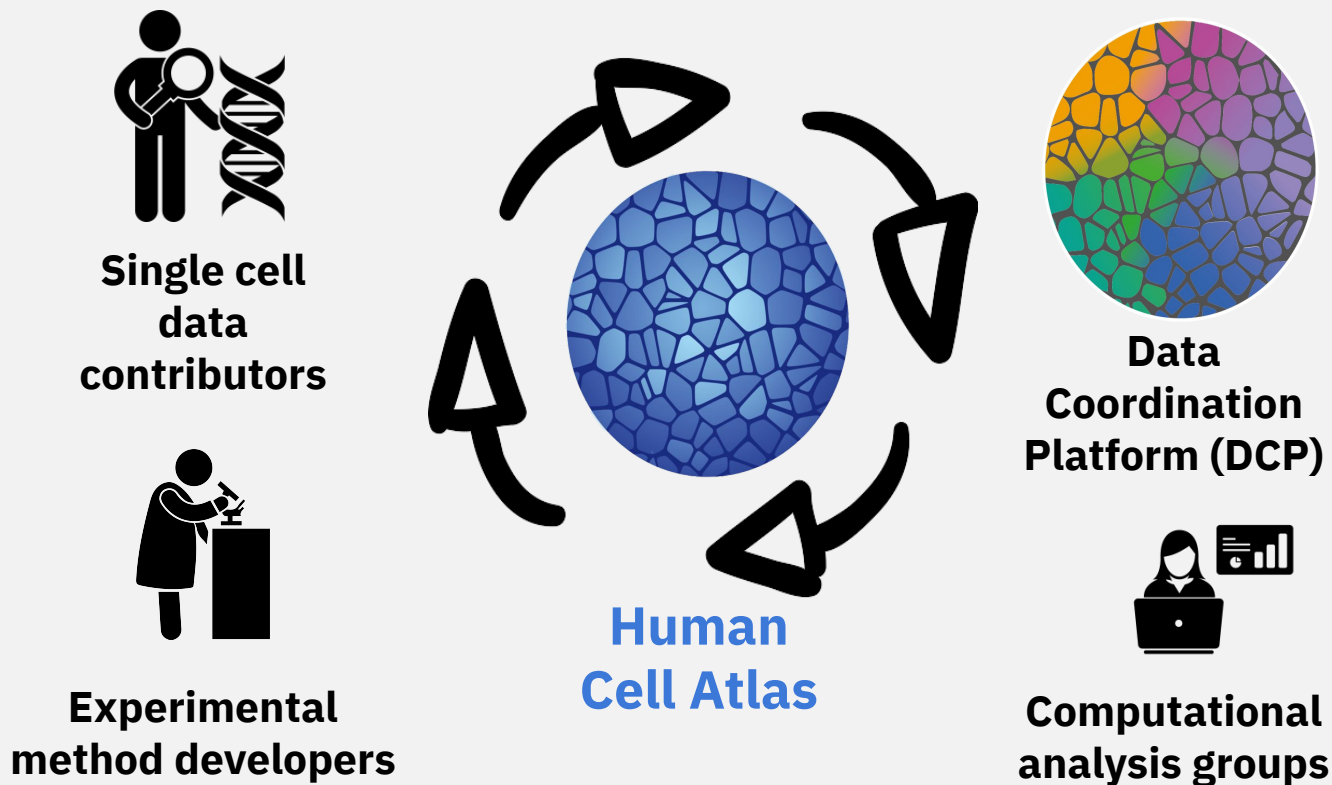


# Cell Ontologies, Annotation & Metadata

**How can we make ontology annotation following standard schemas easy, efficient, and accurate while leaving room for revising and adding to existing ontologies?**



# The Human Cell Atlas Data Coordination Platform (DCP)





# HCA - example of desired metadata

## Donor



NCBI TAXON ID  
GENUS SPECIES  
BIOLOGICAL SEX  
ALIVE AT COLLECTION

KNOWN DISEASE(S)  
ETHNICITY

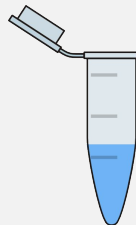
## Specimen from organism



NCBI TAXON ID  
GENUS SPECIES  
ORGAN  
INSDC EXPERIMENT  
ACCESSION

ORGAN PART  
KNOWN DISEASE(S)  
POST-MORTEM INTERVAL  
PRESERVATION METHOD

## Cell Suspension



PERCENT CELL  
VIABILITY  
CELL VIABILITY  
METHOD  
CELL VIABILITY RESULT  
PERCENT NECROTIC  
CELLS

## Sequencing



CELL BARCODE (3)  
INPUT NUCLEIC ACID MOLECULE  
NUCLEIC ACID SOURCE  
LIBRARY CONSTRUCTION METHOD  
END BIAS  
STRAND  
UMI BARCODE (3)  
INSTRUMENT MANUFACTURER AND  
MODEL  
PAIRED END  
SEQUENCING METHOD



# The HCA Metadata Schema



Metadata schemas contain all information required to **understand and interpret** the standard

```
1 "organism_age": {  
2   "description": "Age of organism in Age units measured since birth.",  
3   "pattern": "^[0-9]+\\.|\\.[0-9]*-?[0-9]*\\.|\\.[0-9]*$",  
4   "type": "string",  
5   "user_friendly": "Age",  
6   "example": "20; 45-65",  
7   "guidelines": "Enter either a single value or a range of values. Indicate a range using a hyphen."  
8 },  
9  
10 "organism_age_unit": {  
11   "description": "The unit in which Age is expressed.",  
12   "type": "object",  
13   "isrdf": "http://schor.humanconnectome.org/module/ontology/5.2.5/time_unit_ontology",  
14   "user_friendly": "Age unit",  
15 }
```

*organism\_age*: Age of organism, expressed either as a number or a range (20 | 45-65)  
*organism\_age\_unit*: Unit in which Age is expressed. Only accepts ontologised terms



# The HCA Metadata Schema



The schema can be expanded to validate for ontologies

***Any term accepted under classes***

***UO:0000003 (time unit)***

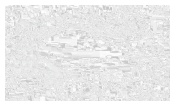
***UO:0000149 (derived time unit)***

Any other term not descending from these 2 is **rejected** in this field

```
26     "ontology": {  
27         "description": "An ontology term identifier in the form prefix:accession",  
28         "type": "string",  
29         "graph_restriction": {  
30             "ontologies": ["obo:uo", "obo:efo"],  
31             "classes": ["UO:0000003", "UO:0000149"],  
32             "relations": ["rdfs:subClassOf"],  
33             "direct": false,  
34             "include_self": false  
35         }
```



# The HCA Metadata Schema

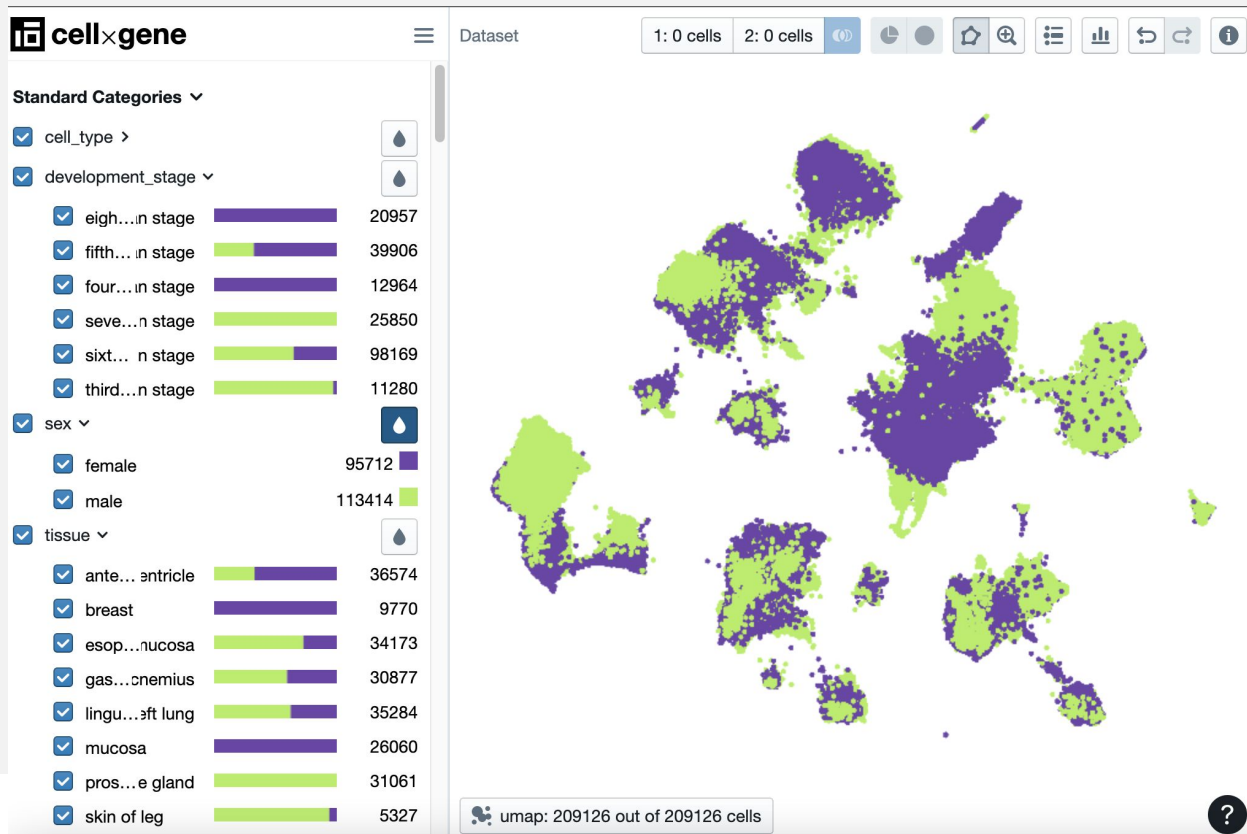


Ethnicity  
Developmental Stage  
Age Unit  
Tissue  
Organ  
Known Diseases  
Library Preparation Methods  
Sequencing Machine  
Cell Cycle  
...





# Cell Ontologies and Annotation





# Cell Ontologies and Annotation



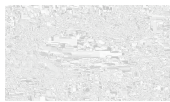
## CellxGene - cell\_type\_ontology\_term\_id

### cell\_type\_ontology\_term\_id

Key	cell_type_ontology_term_id
Annotator	Curator
Value	categorical with <code>str</code> categories. This MUST be a CL term.



# Challenges and Future Development



- Novel Cell Types and adding new terms to CL
- Releasing annotations with data
- Talk to us here or at [wrangler-team@data.humancellatlas.org](mailto:wrangler-team@data.humancellatlas.org)



# Thanks to our partners



## Programs and Funders

- Related and complementary initiatives
- Diverse funded projects across the globe
- Support for central efforts: DCP, meetings, ethics, equity





**Jason Hilton**



Jason Hilton, Stanford U, USA

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 **cellxgene** Data Portal

[cellxgene.cziscience.com](https://cellxgene.cziscience.com)



# CZI Single-Cell Team

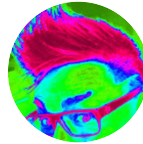
## Product & Design



## Data & Statistics



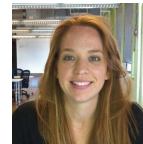
## Engineering



## Science Program



## Clever Canary



# Curation Teams

## Lattice, Stanford Univ.



## HCA DCP, UCSC & EBI



## Sanger



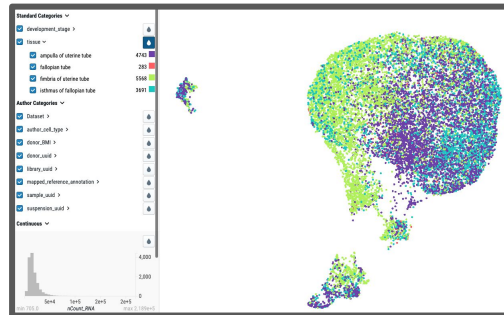
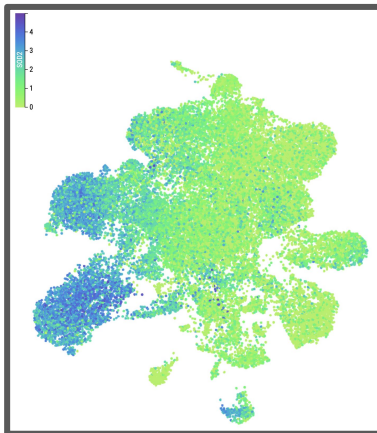
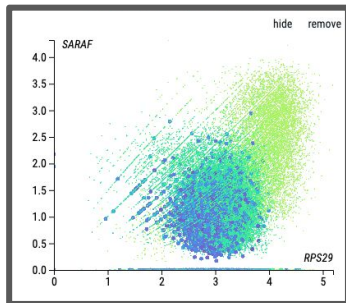
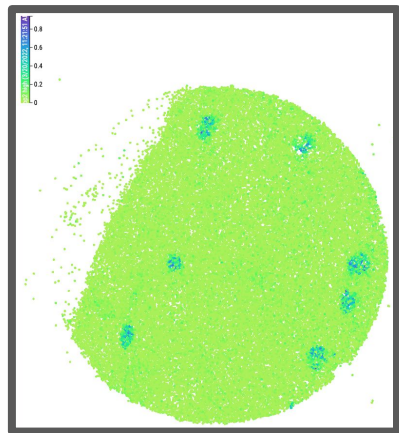


# cellxgene Data Portal

[cellxgene.cziscience.com](https://cellxgene.cziscience.com)

## A. Publish single collection of data

### Visualize and download data





# cellxgene Data Portal

[cellxgene.cziscience.com](https://cellxgene.cziscience.com)

## **A. Publish single collection of data**

Visualize and download data

## **B. Enable cross-collection integration & other reuse cases**

Requires some standardization



# cellxgene Data Portal

Standards: [schema 2.0.0](#)

- All data available as AnnData (.h5ad) & Seurat (.rds)



# cellxgene Data Portal

Standards: [schema 2.0.0](#)

- AnnData & Seurat
- raw counts are required
  - unscaled, pre-normalization counts per cell
  - enable integration without realignment



# cellxgene Data Portal

Standards: [schema 2.0.0](#)

- AnnData & Seurat
- raw counts
- feature metadata
  - Ensembl IDs are required
  - stable identifiers, as opposed to symbols



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Standards: [schema 2.0.0](#)

- AnnData & Seurat
- raw counts
- feature metadata
  - Ensembl IDs are required
  - stable identifiers, as opposed to symbols

## Challenge

**accurately mapping from gene symbols to Ensembl IDs**



# cellxgene Data Portal

Standards: [schema 2.0.0](#)

- AnnData & Seurat
- raw counts
- feature metadata
- observation metadata
  - annotated to the most specific available ontology term

Required field	Ontology
<b>organism</b>	<b>NCBITaxon</b>
<b>donor_id</b>	
<b>development_stage</b>	<b>HsapDv/MmusDv</b>
<b>sex</b>	<b>PATO</b>
<b>ethnicity</b>	
<b>disease</b>	<b>MONDO</b>
<b>tissue</b>	<b>UBERON</b>
<b>cell_type</b>	<b>CL</b>
<b>assay</b>	<b>EFO</b>
<b>observation_type</b>	<b>[cell,nucleus]</b>



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## Challenge

### How to standardize ethnicity

Required field	Ontology
<b>organism</b>	<b>NCBITaxon</b>
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<b>cell_type</b>	<b>CL</b>
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<b>observation_type</b>	<b>[cell,nucleus]</b>



# cellxgene Data Portal

Standards: [schema 2.0.0](#)

- AnnData & Seurat
- raw counts
- feature metadata
- observation metadata
  - annotated to the most specific available ontology term

## Challenge

**How to standardize  
unannotated cells**

## Challenge

**How to standardize ethnicity**

Required field	Ontology
organism	NCBITaxon
donor_id	
development_stage	HsapDv/MmusDv
sex	PATO
ethnicity	
disease	MONDO
tissue	UBERON
cell_type	CL
assay	EFO
observation_type	[cell,nucleus]



# cellxgene Data Portal

## Ontology usage

Group data by higher-level terms

cellxgene Data Portal			
Datasets Collections scExpression <span>BETA</span>			
Filters		Collection	Publication
Assay		SEA-AD: Seattle Alzheimer's Disease Brain Cell Atlas	No publication
Authors			
Cell Type		Single-nucleus cross-tissue molecular reference maps to decipher disease gene	Eraslan et al. (2022) Science
Development Stage			
HOMO SAPIENS		MUS MUSCULUS	
Prenatal (conception–birth)	9	Prenatal	1 al. (2022)
Embryonic human (0–56 days)	3	Embryonic mouse	1 Consortium*
Carnegie (CS1)	0	Thelie stage 1 (TS1)	0
Cleavage (CS2)	0	Cleavage (TS2–3)	0 iScience
Blastula (CS3–5)	0	Blastula (TS4–8)	0
Gastrula (CS6)	0	Gastrula (TS9–10)	0
Neurula (CS7–8)	0	Thelie stage 11 (TS11)	0
Organogenesis (CS9–23)	1	Organogenesis (TS11–22)	1
Fetal (>56 days–birth)	9	Fetal (TS23–26)	1 ell
Immature (0–12 years)	12	Post-partum (Birth+)	15
Newborn human (0–1 month)	3	Immature (0–6 weeks)	0 Nature
Infant (1–23 months)	6	Mature (6+ weeks)	15
Child (2–12 years)	8	Early adulthood (6 weeks–7 months)	15 Nat Commun
Mature (13+ years)	58	Late adulthood (7+ months)	2 2021) Nat
Adolescent (13–19 years)	5		
Human adult (10+ years)	58		
Single cell transcriptional and chromatin accessibility profiling redefine cellular heterogeneity in the adult human kidney		Muto et al. (2021) Nat Commun	
Single cell transcriptomic atlas for adult human		No publication	



# cellxgene Data Portal

## Ontology usage

Organism ▾  
Publication Date ▾  
Sex ▾  
Tissue ▾

Search

fimbria of uterine tube	1
fovea centralis	2
frontal cortex	2
gall bladder	1
gastrocnemius	1
gonadal fat pad	1
heart	3
heart left ventricle	1
heart right ventricle	1
hippocampal field	2

Tabula Sapiens

High Resolution Slide-seqV2 Spatial Transcriptomics Enables Discovery of Disease-Associated Cell Neighborhoods and Pathways

Blockade prevents injury to the pulmonary gas exchange surface in

Sequencing of follicular

COVID-19 defines hallmarks of and specificity

Immune responses to SARS-CoV-2 in children and adults

Identified mRNA signals across human

Mapping the temporal and spatial dynamics of the human endometrium in vivo and in vitro

Group data by higher-level terms

Grouping tissue

“system” and/or “organ”

**Challenge**

**How to group by  
higher-level terms**

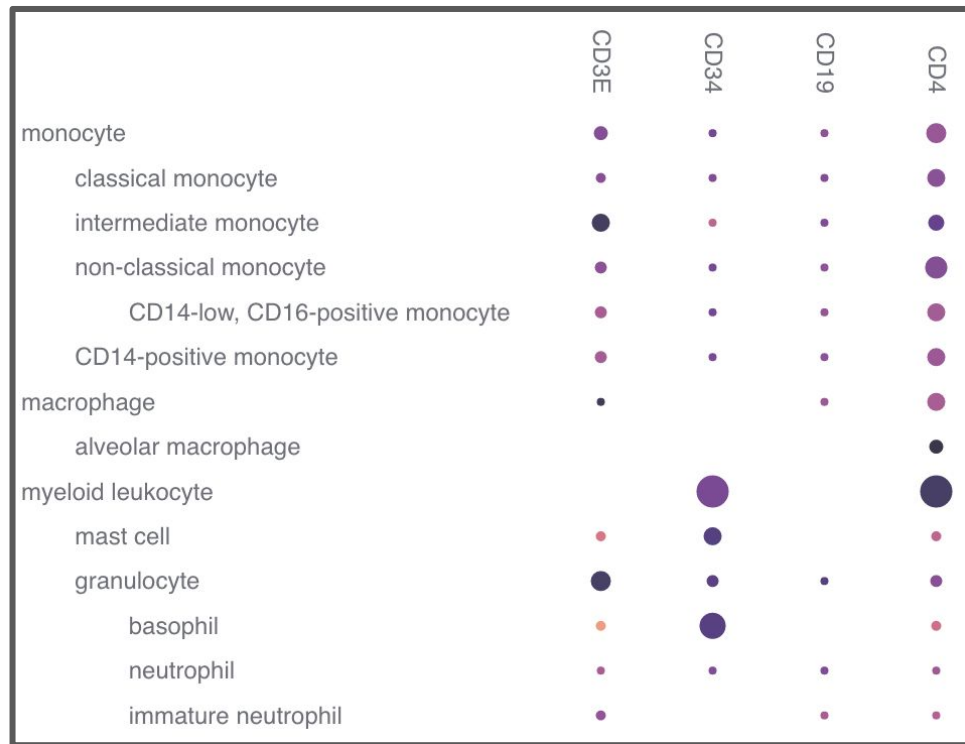


# cellxgene Data Portal

## Ontology usage

[Datasets](#)[Collections](#)[scExpression](#)[BETA](#)

## Ontology-aware organization of per-cell expression





# cellxgene Data Portal

## Challenges

- Mapping Ensembl IDs from gene symbols
- Standardizing ethnicity
- Standardizing unannotated cells
- Understanding how users want to find & filter data

**THANK YOU!**



**Evan Biederstedt**



# Evan Biederstedt, Harvard Medical School, USA

— — —

<https://speakerdeck.com/evanbiederstedt/hca-general-meeting-2022-cell-annotation-platform>

<https://rc1.celltype.info> - Demo time! (**Update:** on YouTube soon)

9:00 AM



9:00 AM - 10:30 AM GMT +2 / 3:00 AM - 4:30 AM Your local time (1 Hour, 30 Min)

## Plenary Session 4 - Opportunities and Challenges Using and Accessing Data

Palais Niederosterreich, Main Hall

Session Chair, Leyla Tarhan

1. Nile Gehlbach: "Atlas User Interface to Accelerate Data Access"



**Irene Papatheodorou**

Team Leader  
EMBL-EBI  
Speaker



**Malte Luecken**

Postdoctoral Researcher  
Helmholtz Munich  
Speaker



**Evan Biederstedt**

Research Scientist  
Harvard Medical School  
Speaker



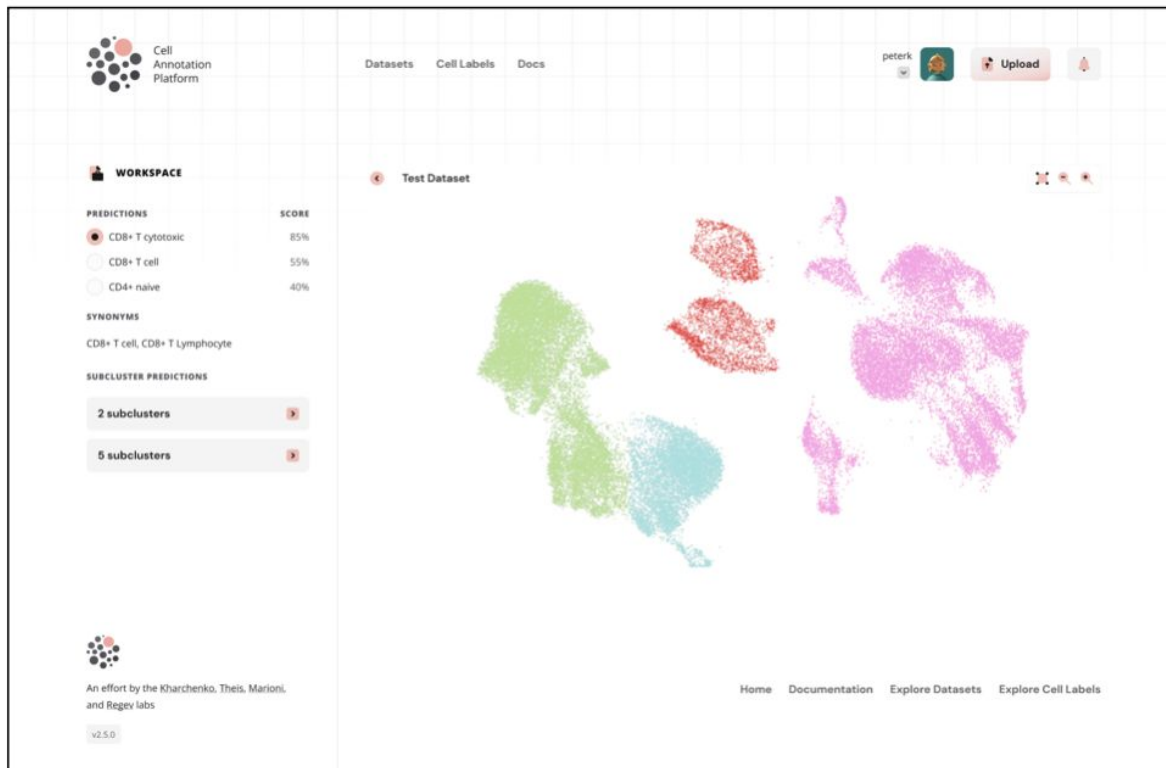
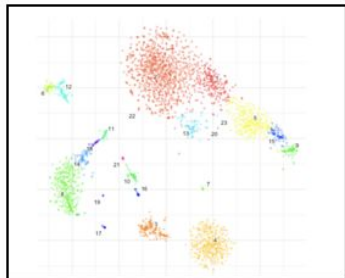


# Annotation Suggestions in Real-time

## Text-based



## Molecular-based





# Evan Biederstedt, Harvard Medical School, USA

---



## User Feedback Request

- **Prioritize Future Features**
  - Annotation Feedback?
  - Community Ratings?
  - Evidence?
  - Contrast/Compare Annotation A vs B?
  - Specific UI Requests?
- **Demos & User Feedback Surveys**



**Talk to us!**



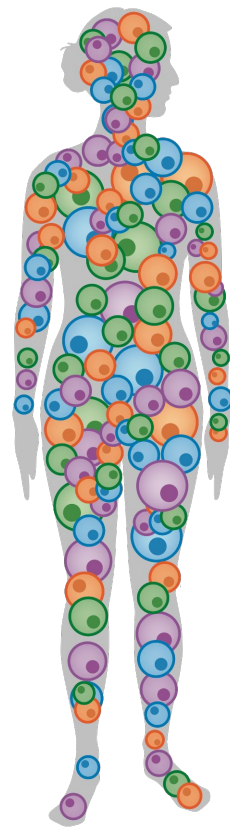
**Angela Pisco**



Angela Pisco, CZ Biohub, USA

---

## Tools for Automating Cell Type Calling



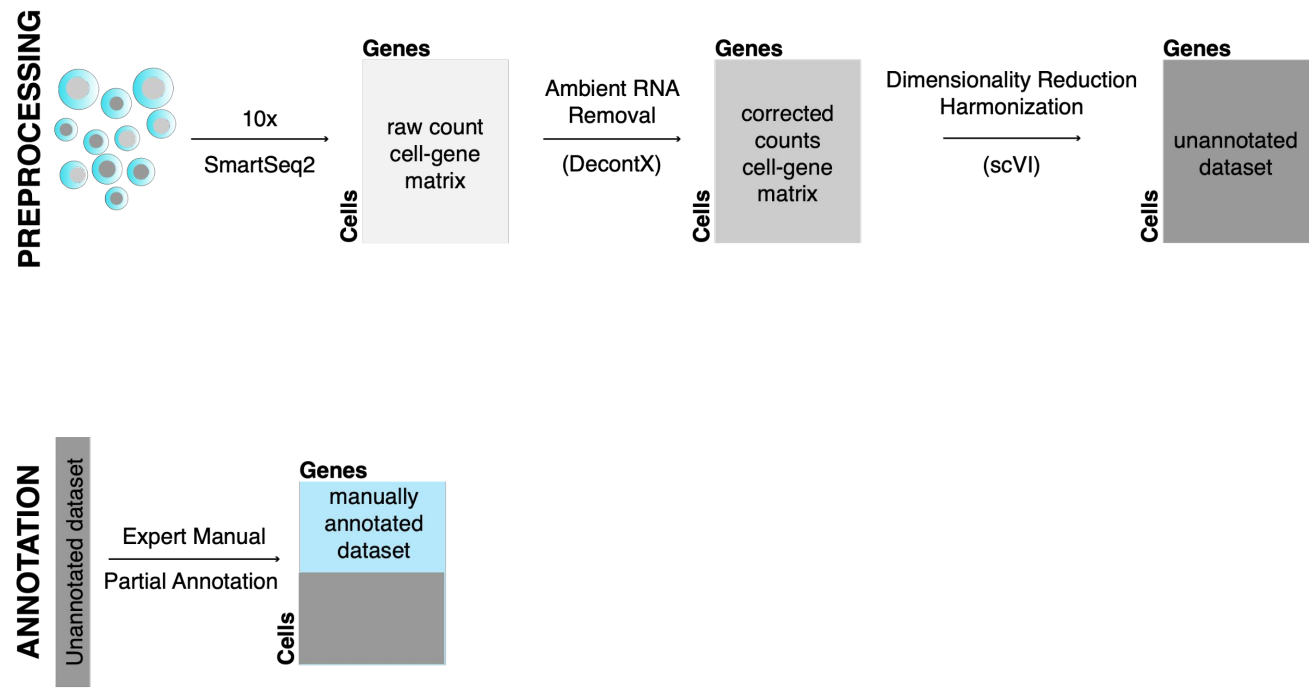
 [angela.pisco@czbiohub.org](mailto:angela.pisco@czbiohub.org)

 [@drAOPisco](https://twitter.com/adrAOPisco)

 [@aopisco](https://github.com/aopisco)



# CELL TYPE ANNOTATION WHEN BUILDING AN ATLAS






# CELL TYPE ANNOTATION TOOLS (I)

Automated cell  
type annotation

Article | [Published: 19 October 2020](#)

## MARS: discovering novel cell types across heterogeneous single-cell experiments

[Maria Brbić](#), [Marinka Zitnik](#), [Sheng Wang](#), [Angela O. Pisco](#), [Russ B. Altman](#), [Spyros Darmanis](#) & [Jure Leskovec](#) 

*Nature Methods* (2020) | [Cite this article](#)

Article | [Open Access](#) | [Published: 21 September 2021](#)

## Leveraging the Cell Ontology to classify unseen cell types

[Sheng Wang](#), [Angela Oliveira Pisco](#) , [Aaron McGeever](#), [Maria Brbic](#), [Marinka Zitnik](#), [Spyros Darmanis](#), [Jure Leskovec](#), [Jim Karkanias](#) & [Russ B. Altman](#) 

*Nature Communications* **12**, Article number: 5556 (2021) | [Cite this article](#)



# CELL TYPE ANNOTATION TOOLS (II)

Article | [Published: 19 October 2020](#)

## MARS: discovering novel cell types across heterogeneous single-cell experiments

Maria Brbić, Marinka Zitnik, Sheng Wang, Angela O. Pisco, Russ B. Altman, Spyros Darmanis & Jure Leskovec [✉](#)

*Nature Methods* (2020) | [Cite this article](#)

Article | [Open Access](#) | [Published: 21 September 2021](#)

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*Nature Communications* **12**, Article number: 5556 (2021) | [Cite this article](#)



ANNOTATION

Unannotated dataset

Expert Manual  
Partial Annotation

Cells

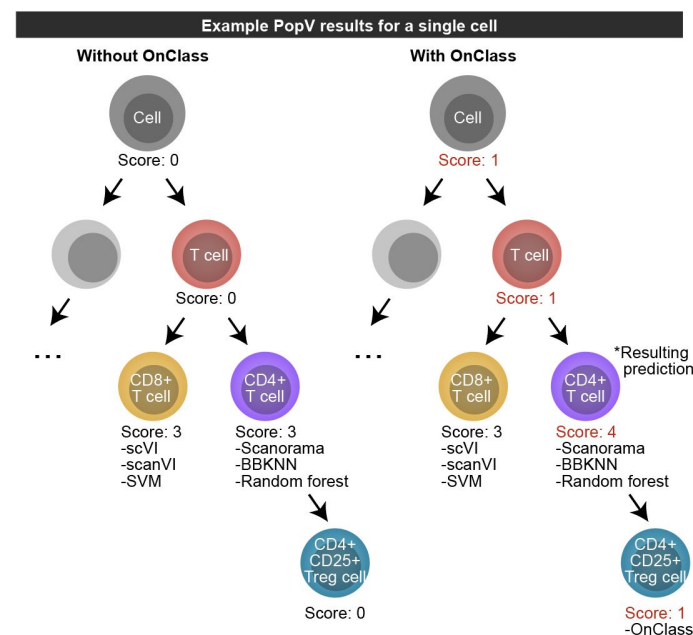
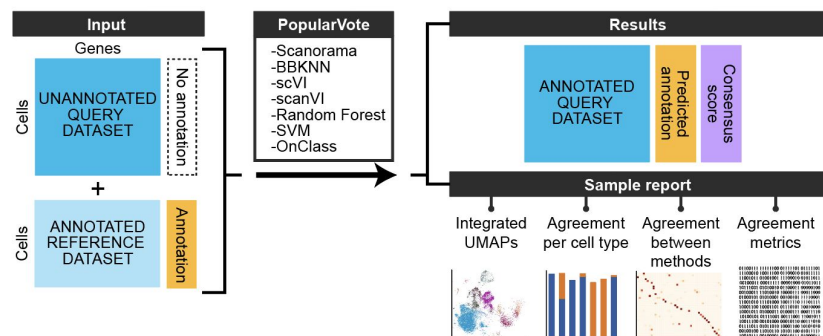
Genes  
manually  
annotated  
dataset

Cells

Genes  
computer  
automated  
annotated  
dataset

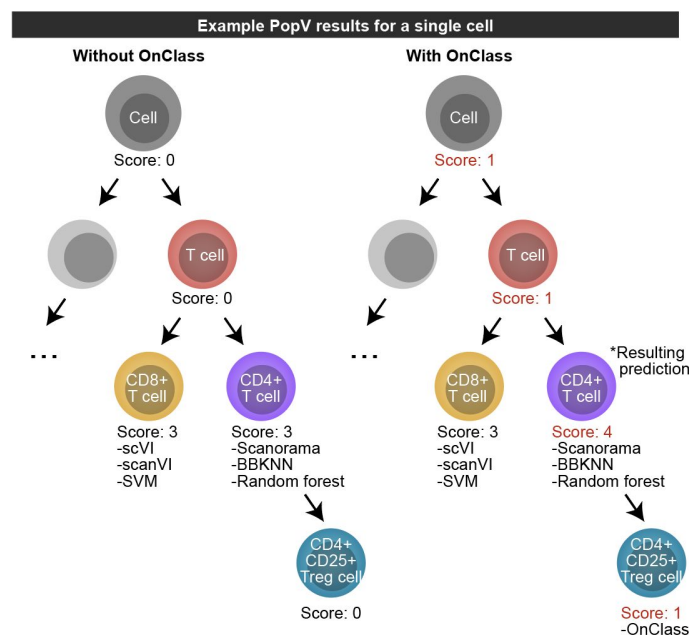
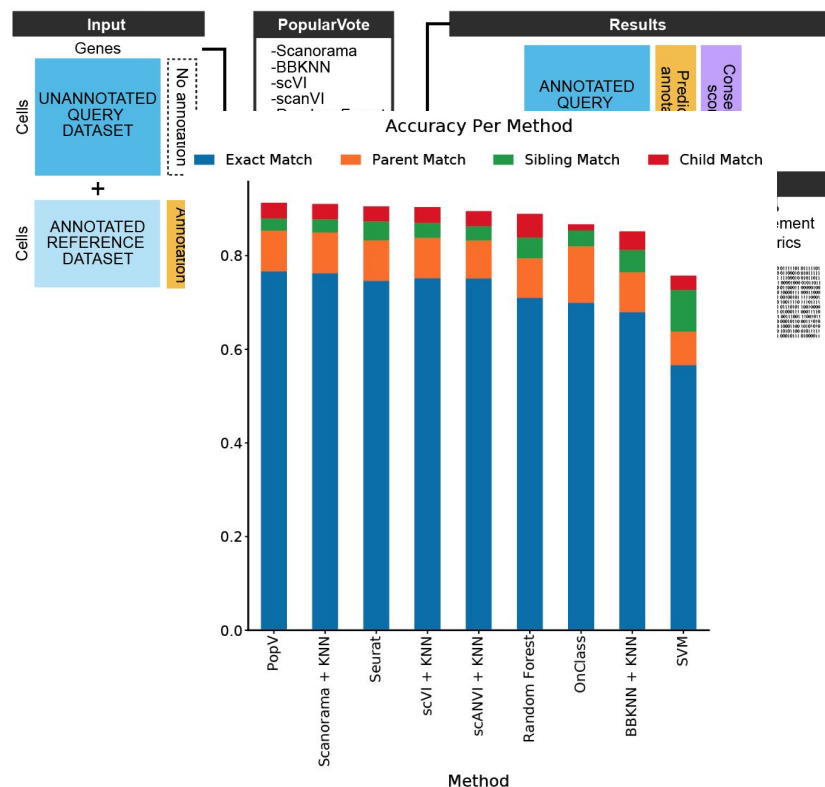


# CELL TYPE ANNOTATION TOOLS (III)



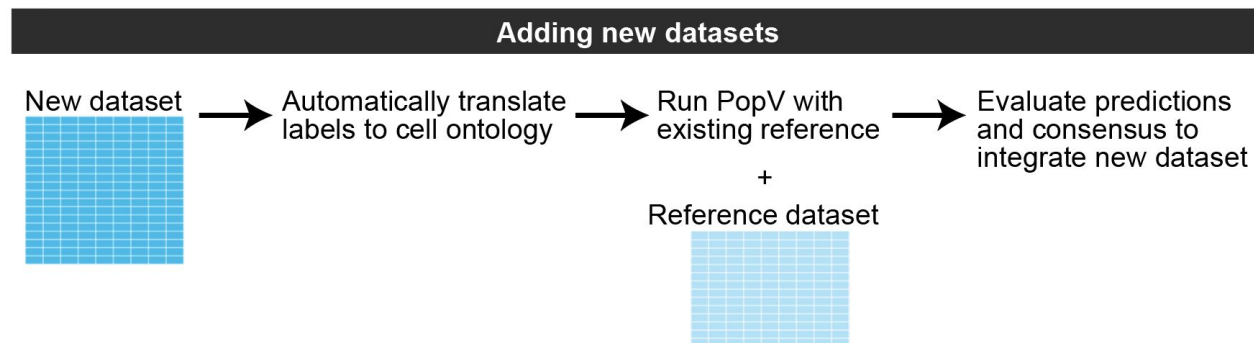


# CELL TYPE ANNOTATION TOOLS (III)





# CELL TYPE ANNOTATION TOOLS (IV)





# CELL TYPE ANNOTATION TOOLS (V)

Automated cell  
type annotation

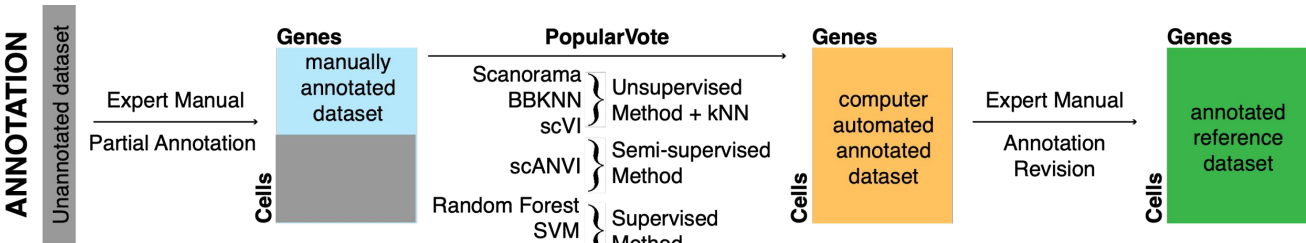
Article | Published: 19 October 2020

## MARS: discovering novel cell types across heterogeneous single-cell experiments

Maria Brbić, Marinka Zitnik, Sheng Wang, Angela O. Pisco, Russ B. Altman, Spyros Darmanis & Jure Leskovec

Nature Methods (2020) | Cite this article

Using the Tabula datasets as general  
reference for annotations



Article | Open Access | Published: 21 September 2021

## Leveraging the Cell Ontology to classify unseen cell types

Sheng Wang, Angela Oliveira Pisco, Aaron McGeever, Maria Brbic, Marinka Zitnik, Spyros Darmanis, Jure Leskovec, Jim Karkanias & Russ B. Altman

Nature Communications 12, Article number: 5556 (2021) | Cite this article



# MARKER GENES FOR CELL TYPES

## OnClass

Single cell typing based on cell ontology.

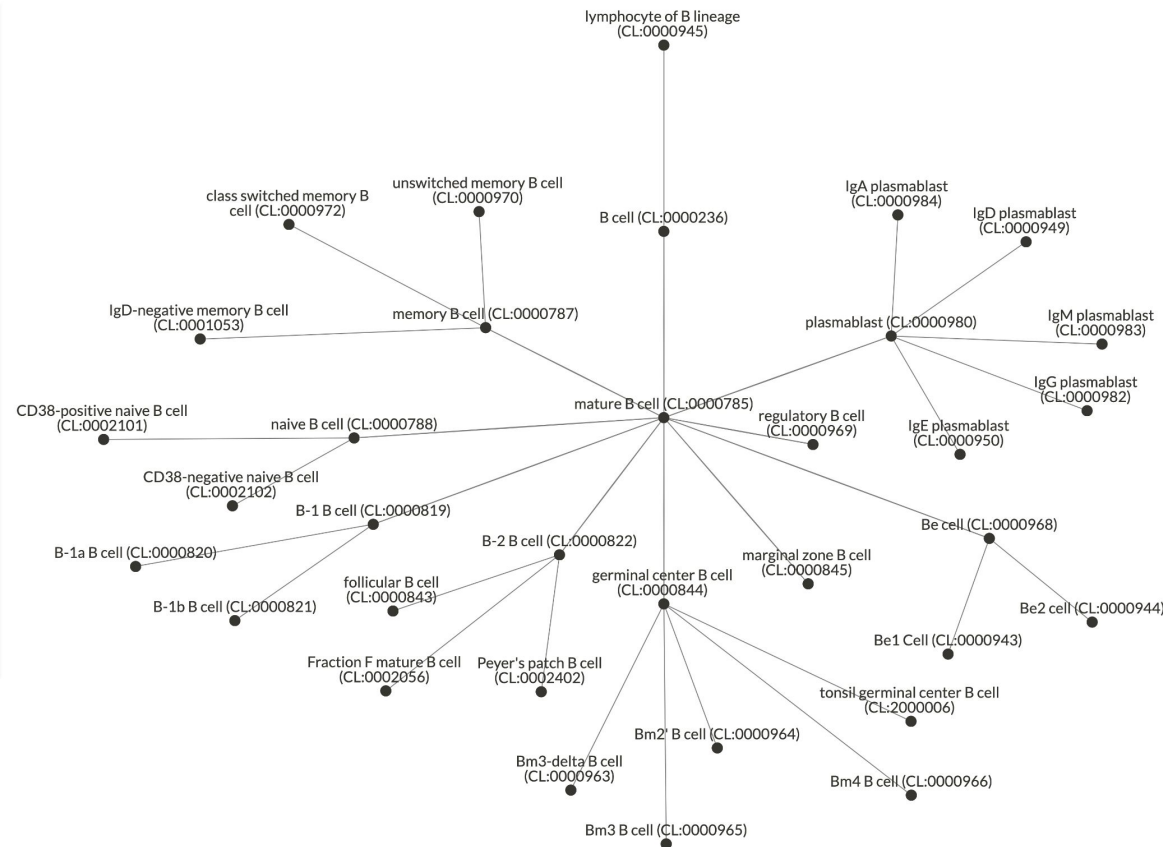
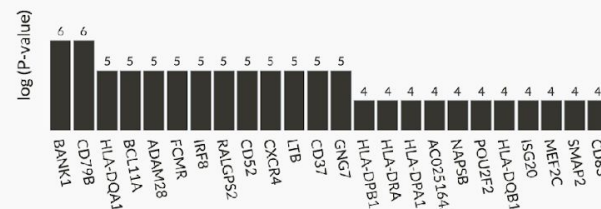
[Paper](#) [Read the Docs](#) [GitHub](#)

Choose a cell type:

mature B cell (CL:0000785)

### mature B cell (CL:0000785)

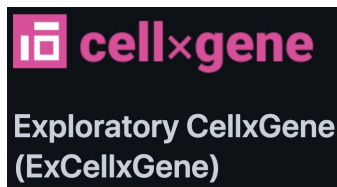
A B cell that is mature, having left the bone marrow. Initially, these cells are IgM-positive and IgD-positive, and they can be activated by antigen. [GOC:add, GOC:dsd, ISBN:0781735149]



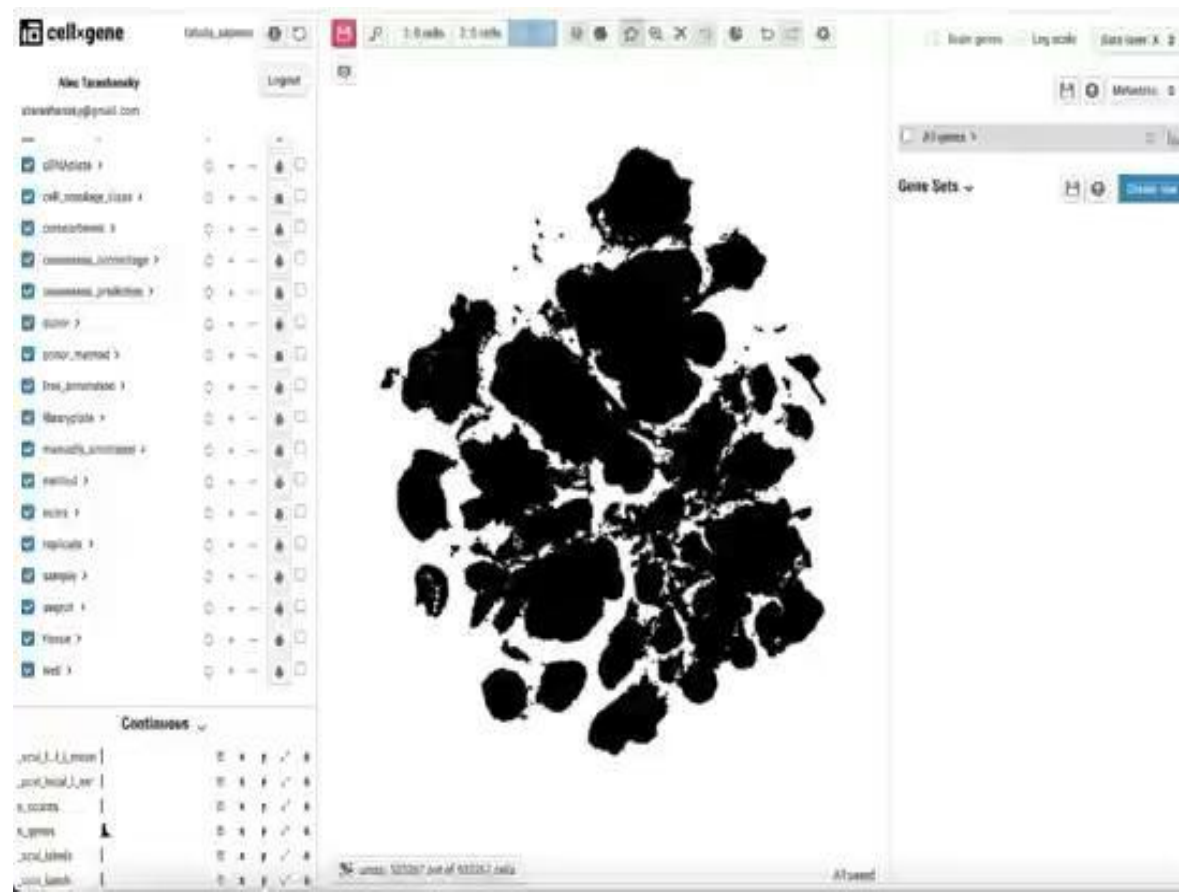




## EXCELLXGENE



- Lidar
- Differential gene expression on the fly
- Nested embeddings
- Leiden clustering
- Re-embeddding
- Sankey diagrams





# THANK YOU!

**The Tabula Muris consortium**

**The Tabula Sapiens consortium**

**The Tabula Microcebus consortium**

**The Fly Cell Atlas consortium**

**The Covid Tissue Atlas consortium**

**CZBiohub Data Science Team**

**CZBiohub Genomics Team**



**We are  
hiring!**



Ahmad Salehi  
Ravi Ponnusmi

We express our gratitude and thanks to donor WEM and his family, as well as all of the anonymous organ and tissue donors and their families for giving both the gift of life and the gift of knowledge by their generous donations.



**CZI**  
Collin Megill  
Max Lombardo  
Ambrose Carr  
Jenn Tang  
Tiago Carvalho



**Fabian Theis**



# Fabian Theis, Helmholtz Munich, Germany

— — —

Learning and using gene set ontologies in single cell genomics



# Data management & annotation with sfaira

## Data curation:

- one loader per study, publicly maintained
- easy to contribute, easy to extend
- allow generation of streamlined .h5ad files, e.g: *cellxgene*

## Data collection management:

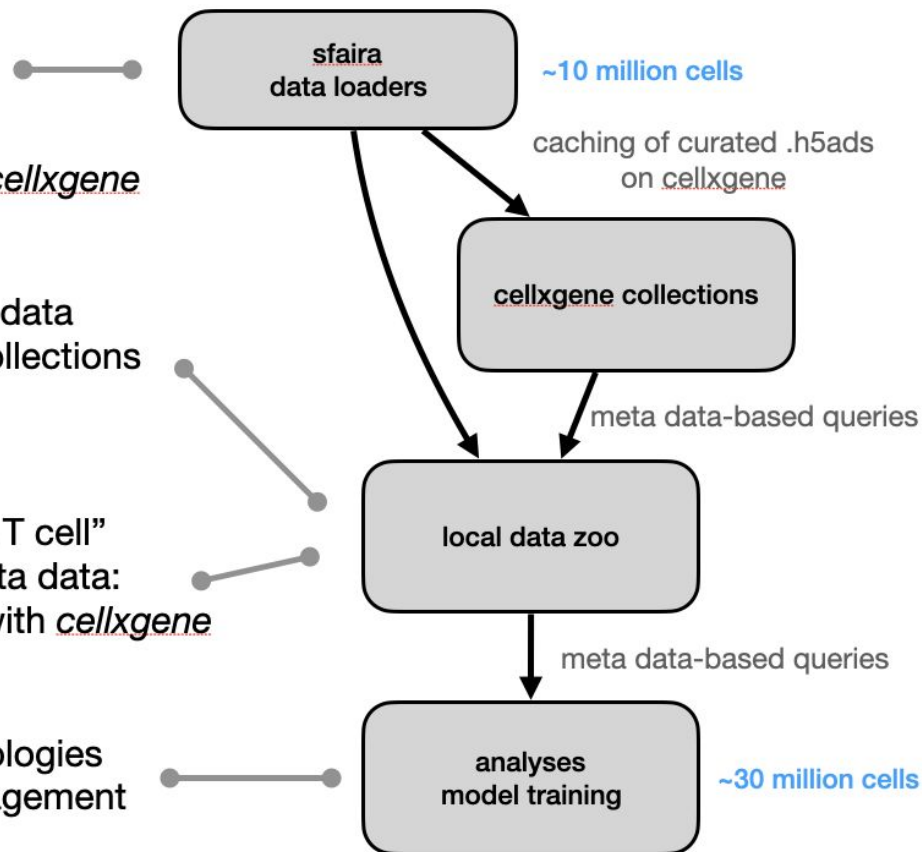
- select datasets from a collection based on meta data
- interface local and public (e.g. *cellxgene*) data collections
- prepare data for model training

## Leverage meta data:

- ontology-based queries to data: subset to “is a T cell”
- uses publicly curated ontologies for all major meta data: cell type, tissue, organism, disease, ..., synced with *cellxgene*

## Models:

- syntax for feature and label space based on ontologies
- > seamless integration with data collection management





# Case study: Preparing HLCA for cellxgene

## Data curation:

- map meta data annotation to ontologies
  - format fields in AnnData object to satisfy cellxgene requirements
- effective time for writing data loader: ~ 1-2 hours

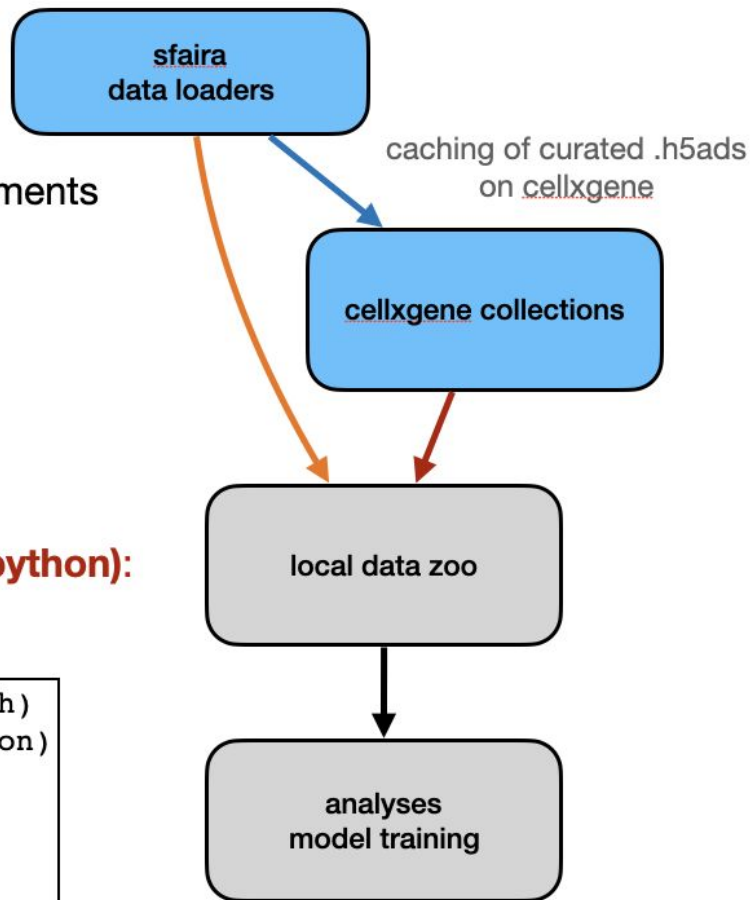
## Data collection management:

Load HLCA AnnData via either mechanism:

- via sfaira data loader:
  - load non-curated (as published in paper) AnnData
  - load curated AnnData (any curation format)
- via cellxgene (**website**) or sfaira interface of cellxgene (**in python**):
  - load curated AnnData (cellxgene format)

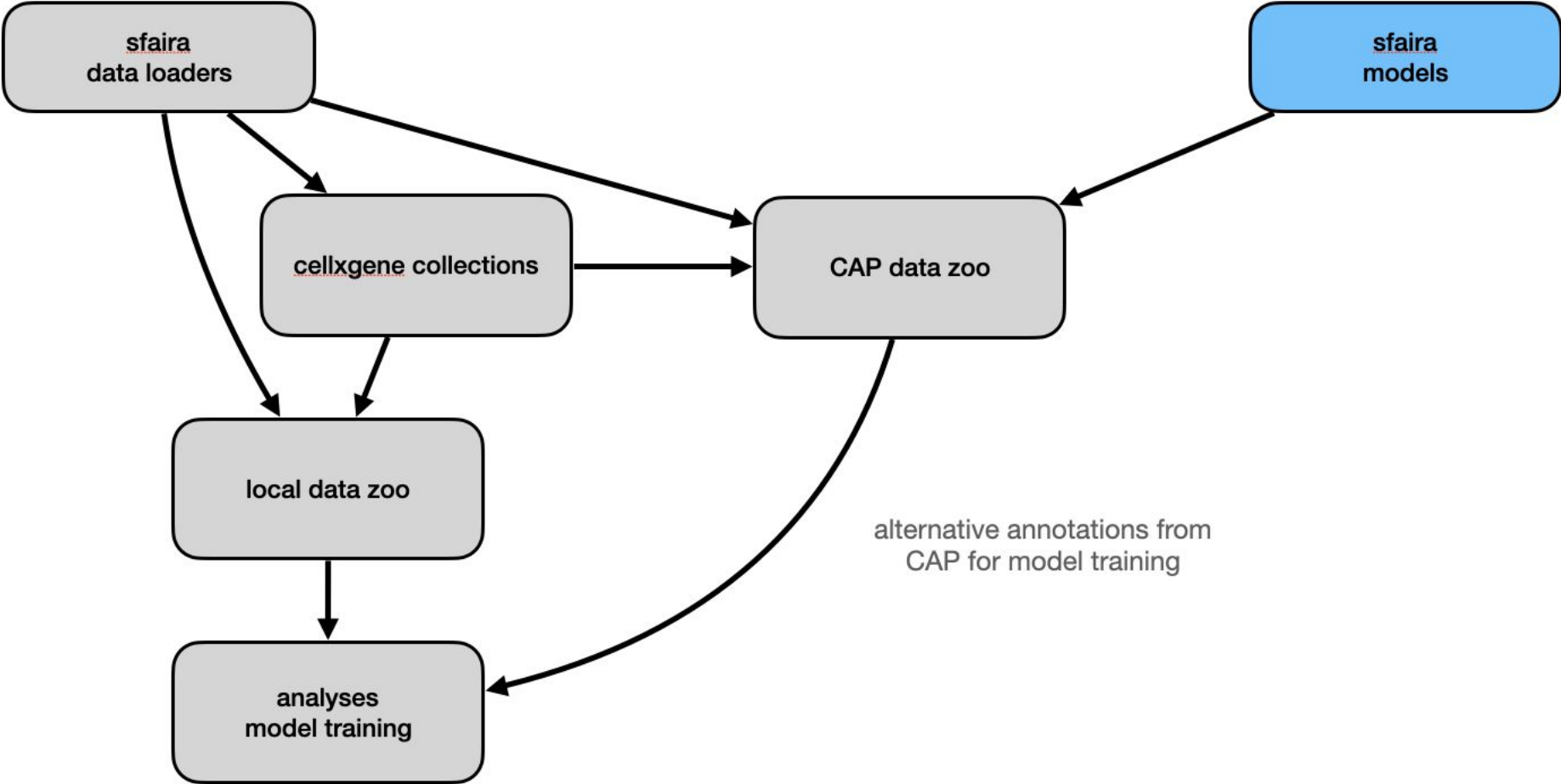
sfaira interface  
of cellxgene

```
dsg = DatasetSuperGroupDatabases(data_path=cache_path)
dsg.subset(key="collection_id", values=HLCA_collection)
dsg.download()
dsg.load()
adatas_hlca = dsg.adata_ls
```



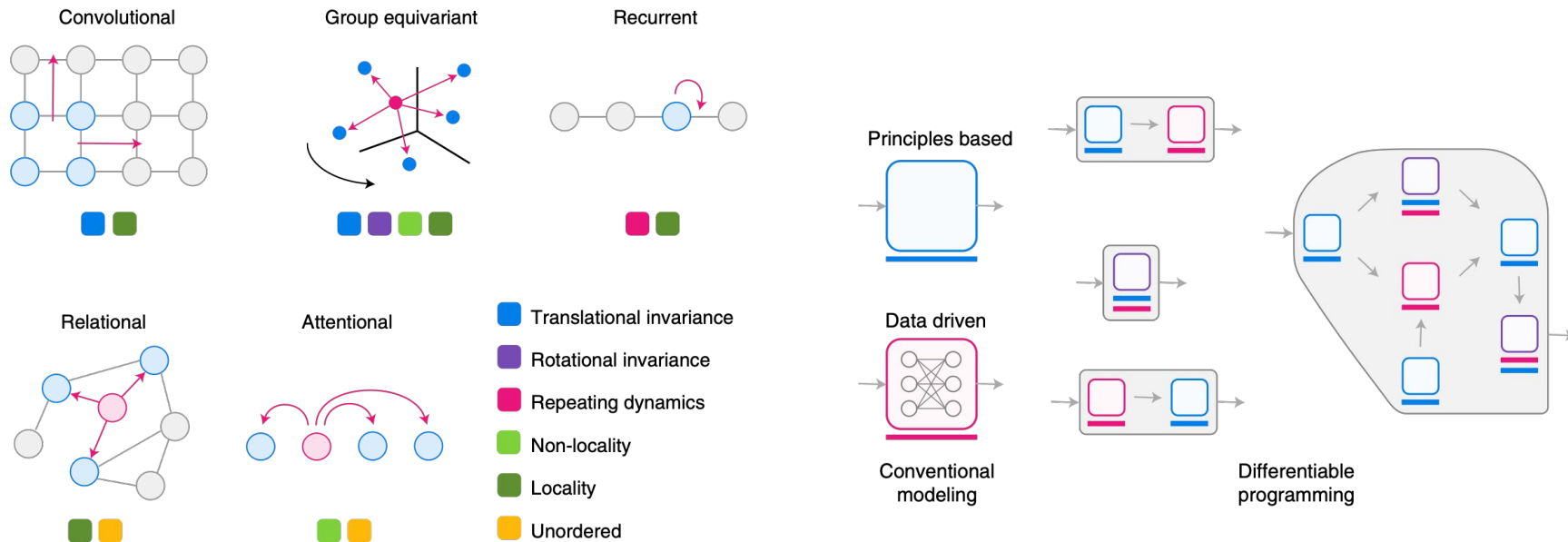


# Sfaira, CAP, cellxgene





# Outlook: differential biology - deep learning for modeling molecular mechanisms

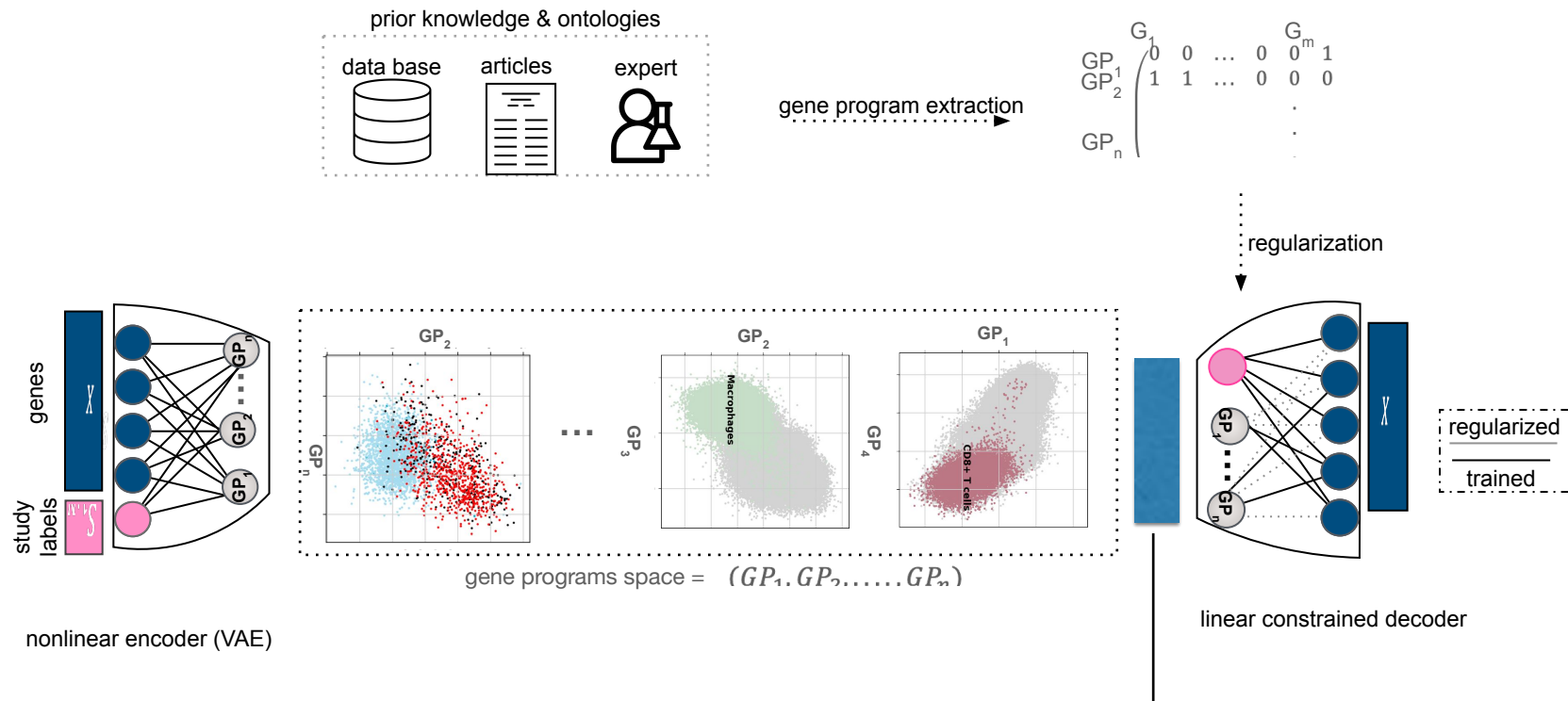


network primitives and invariances

→ questions: reuse primitives? add constraints?

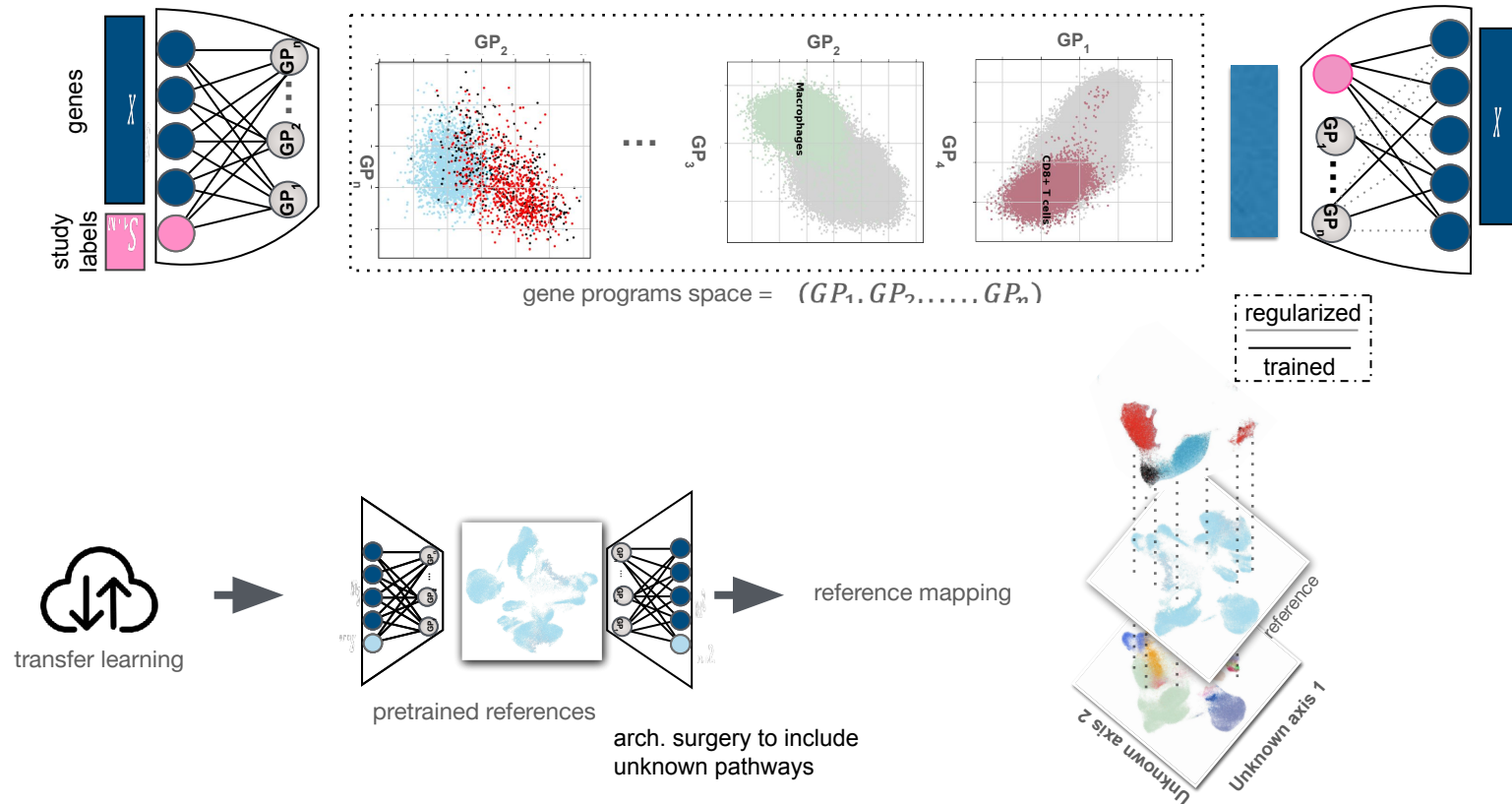


# Interpretable atlas querying: explainable programmable mapper (expiMap)





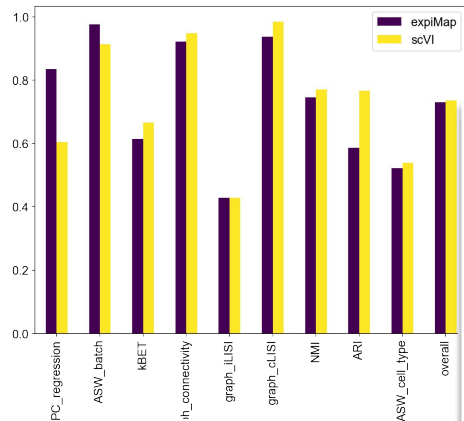
# Interpretable atlas querying: explainable programmable mapper (expiMap)





# expiMap does not loose expressiveness versus nonlinear models

blood (8 batches)

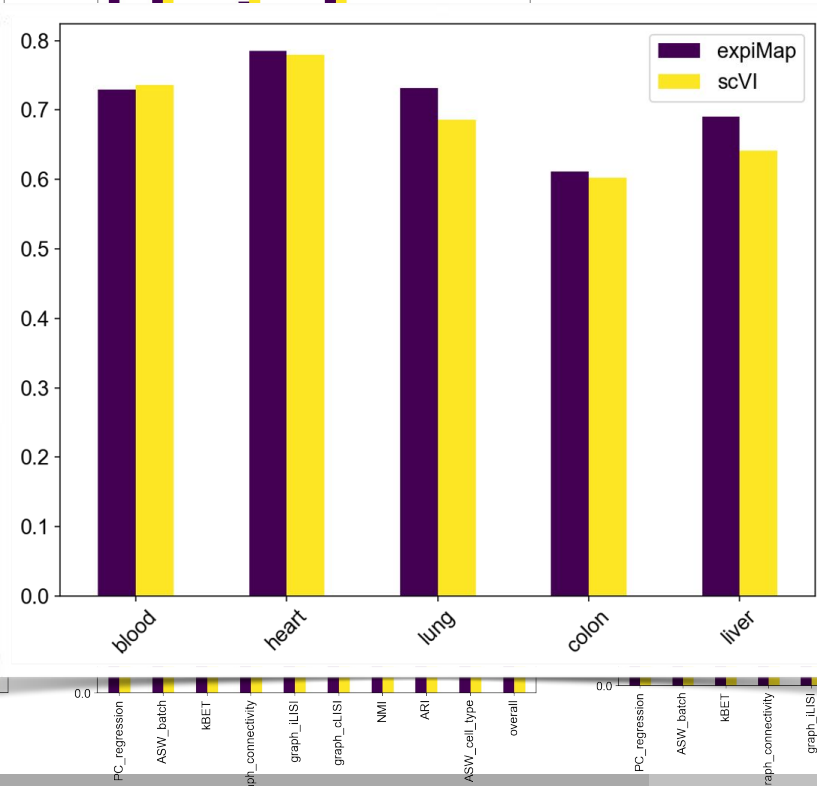
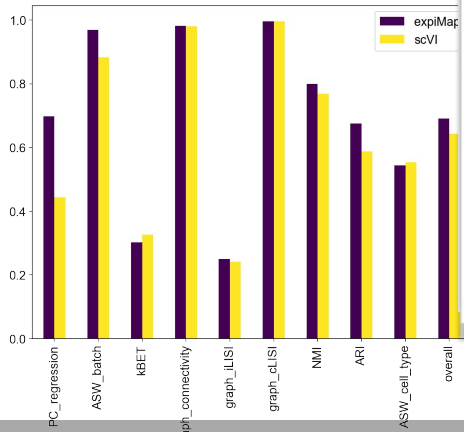


colon (12 batches)



**metrics:** single-cell integration benchmark  
([github.com/theislab/scIB](https://github.com/theislab/scIB))  
(Luecken et al, Nat Meth in press)

liver (14 batches)



batches)



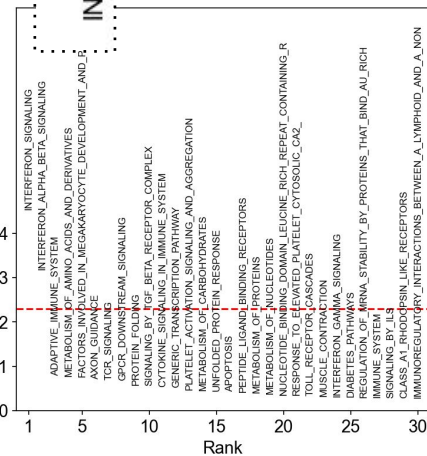
source: sfaira multi-study organ batches  
Fischer, Dony et al, *Genome Biology* 2021



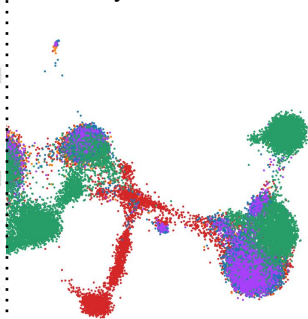
# expmap approximates latent space structure in interpretable fashion

Full GPs

Absolute log bayes factors

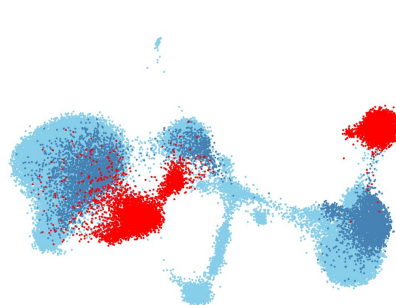


study



10X  
Freitag  
Kang  
Oetjen  
Sun

data source



control (query)  
control (reference)  
stimulated (query)

cell type



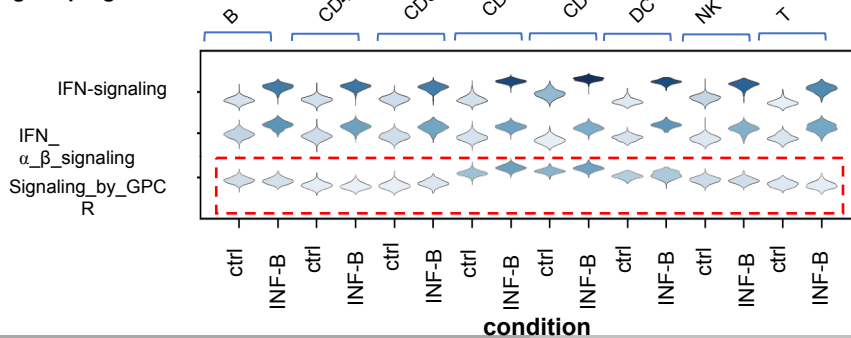
B  
CD4+ T cells  
CD8+ T cells  
CD10+ B cells  
CD14+ Monocytes  
CD16+ Monocytes  
Erythrocytes  
Erythroid progenitors  
HSPCs  
Megakaryocyte progenitors  
Monocyte progenitors  
Monocyte-derived dendritic cells  
NK cells  
NKT cells  
Plasma cells  
Plasmacytoid dendritic cells  
T

Median expression  
in group



cell types

gene programmes





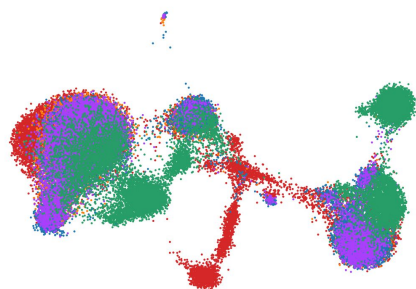
# expiMap approximates latent space structure in interpretable fashion

study

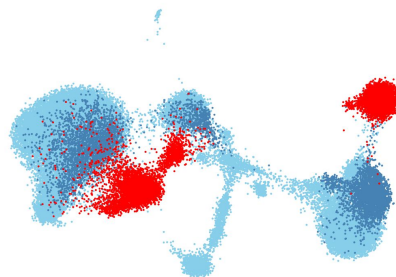
data source

cell type

Full GPs



- 10X
- Freytag
- Kang
- Oetjen
- Sun

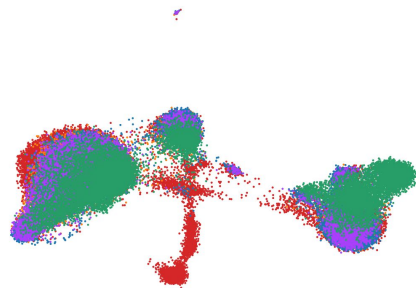


- control (query)
- control (reference)
- stimulated (query)

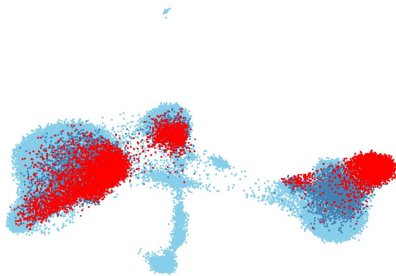


- B
- CD4+ T cells
- CD8+ T cells
- CD10+ B cells
- CD14+ Monocytes
- CD16+ Monocytes
- DC
- Erythrocytes
- Erythroid progenitors
- HSPCs
- Megakaryocyte progenitors
- Monocyte progenitors
- Monocyte-derived dendritic cells
- NK cells
- NKT cells
- Plasma cells
- Plasmacytoid dendritic cells
- T

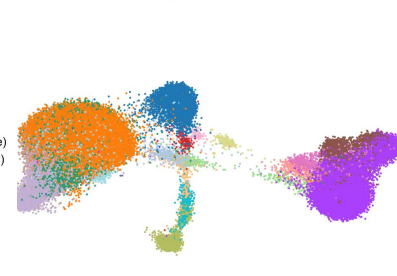
Top 5 IFN- $\beta$  removed



- 10X
- Freytag
- Kang
- Oetjen
- Sun



- control (query)
- control (reference)
- stimulated (query)

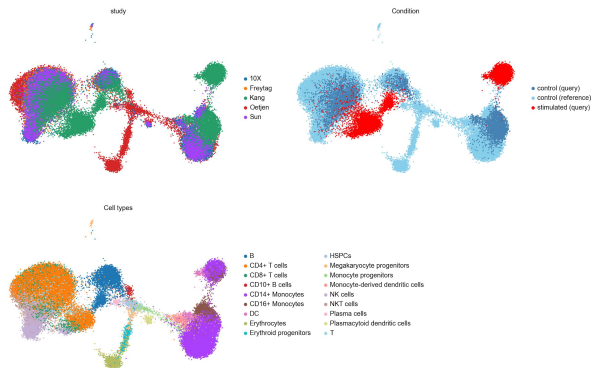


- B
- CD4+ T cells
- CD8+ T cells
- CD10+ B cells
- CD14+ Monocytes
- CD16+ Monocytes
- DC
- Erythrocytes
- Erythroid progenitors
- HSPCs
- Megakaryocyte progenitors
- Monocyte progenitors
- Monocyte-derived dendritic cells
- NK cells
- NKT cells
- Plasma cells
- Plasmacytoid dendritic cells
- T

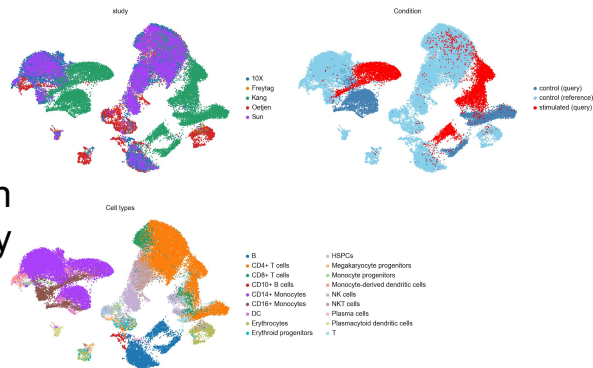


# expiMap integrates cross experiment data while retaining perturbation effect

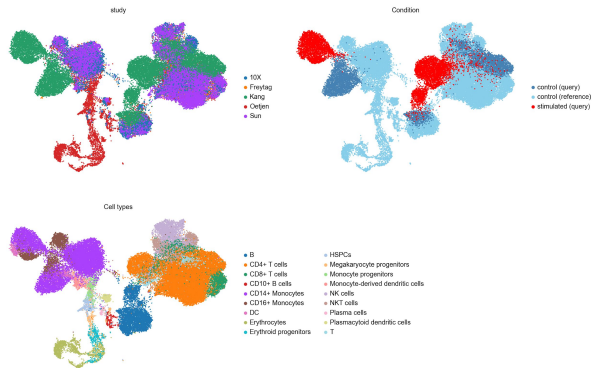
expiMa  
p



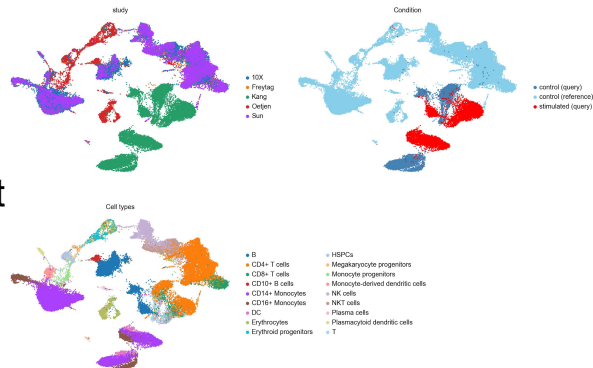
Symphon  
y



scVI



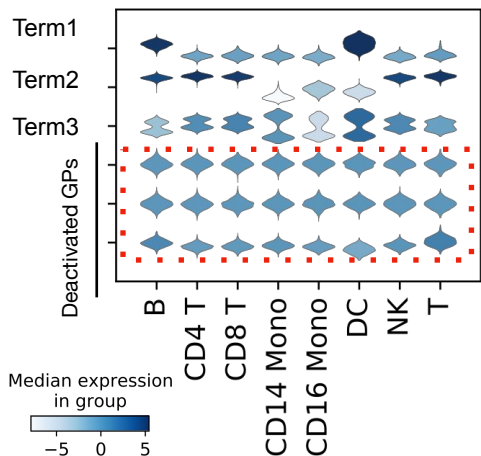
Seurat



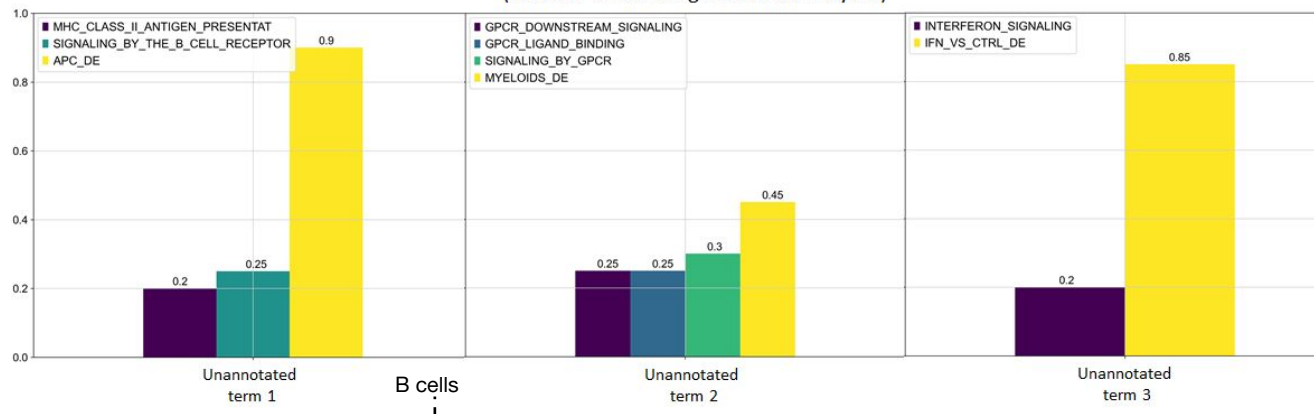


# Learning new interpretable programs

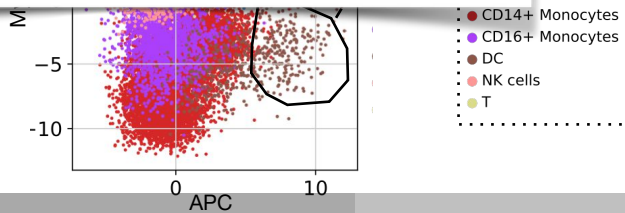
removed B cells & related pathways (incl Inf) from reference  
 -> challenge model to find them during query mapping



Intersection of top 20 genes with different gene sets.  
 (number of shared genes divided by 20)



**ongoing:** transferring learned interpretable embedding from PBMCs to Covid samples helps identifying differential communication pathway during moderate and severe COVID-19



**source**

- control (query)
- control (reference)
- INF-Beta

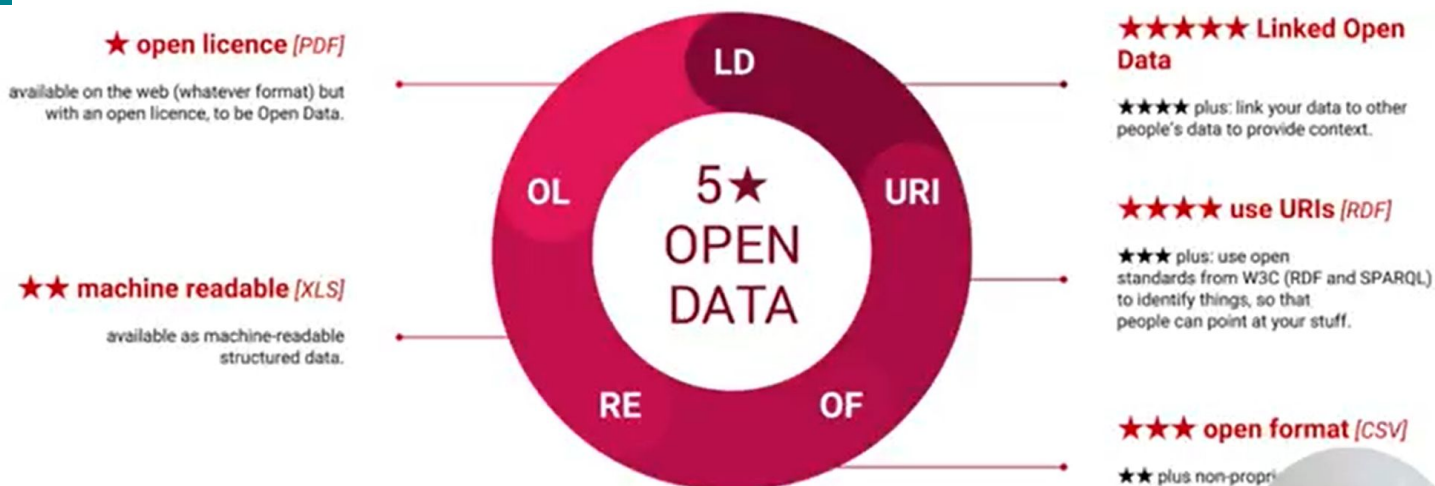


**Bruce W. Herr II**

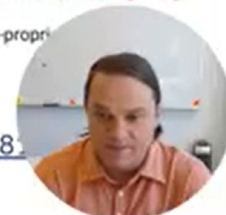


# Bruce W. Herr II, Indiana University, USA

[Slides](#) | [Video](#)



By Florian Thiery - Own work, CC BY 4.0, <https://commons.wikimedia.org/w/index.php?curid=782>





# Discussion



# Discussion Topics

— — —

1. How can we **extend and improve ontologies** as our knowledge grows leveraging expert input, experimental data and feedback from different atlas efforts?
2. How can we **make ontology annotation, following standard schemas, easy**, efficient, and accurate while leaving room for revising and adding to existing ontologies?
3. How can we enable downstream users to take advantage of ontology structure and content in **analysis, visualization and machine learning pipelines/applications**?
4. How can improved annotation with ontologies and the use of linked open data (LOD) help us to interlink atlas data and from multiple consortia and **construct more integrated, coherent, and queryable atlases**?



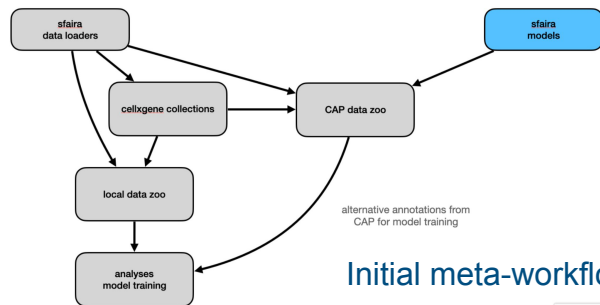
# Report Back



# Cell Ontology, Annotation & Metadata Breakout - Report

## Next Steps

- **Agree on Portal Workflow(s):** Where should users go to annotate their data?
  - How should data flow between portals?
  - Should we encourage annotation to integrated/consensus/cross modal analysis?
- **Agree on** Versioning for ATLAS data and portals
  - So that we can track change as data moves.
- **Agree on** Dataflows for new cell type claims
  - Provisional Cell Ontology (semi-automated)
  - Cell Ontology (curated)



Initial meta-workflow by Fabian Theis

## Other Recommendations

- Retain user free text annotations in addition to ontology annotations
- Include confidence scores with cell type projections
- Cell annotation: Evidence for cell types is needed - but how can we record it well
  - Markers? Projection algorithms? Free text?
- Integrating with spatial data
  - We need reference atlases for anatomical regions.
  - Cell segmentation: Collect gold standards for anatomical and cell segmentations. Run algorithm comparisons, e.g., via Kaggle (HuBMAP+HPA “[Hacking the Human Body](#)”)

[https://cns-iu.github.io/workshops/2022-06-27\\_human\\_cell\\_atlas](https://cns-iu.github.io/workshops/2022-06-27_human_cell_atlas)